



Sulforaphane - role in aging and neurodegeneration

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Abstract In the last several years, numerous molecules derived from plants and vegetables have been tested for their antioxidant, anti-inflammatory, and anti-aging properties. One of them is sulforaphane (SFN), an isothiocyanate present in cruciferous vegetables. SFN activates the antioxidant and anti-inflammatory responses by inducing Nrf2 pathway and inhibiting NF- κ B. It also has an epigenetic effect by inhibiting HDAC and DNA methyltransferases and modifies mitochondrial dynamics. Moreover, SFN preserves proteome homeostasis (proteostasis) by activating the proteasome, which has been shown to lead to increased cellular lifespan and prevent neurodegeneration. In this review, we describe some of the molecular and physical characteristics of SFN, its mechanisms of action, and the effects that SFN treatment induces in order to discuss its relevance as a “miraculous” drug to prevent aging and neurodegeneration.

Keywords Proteasome · Oxidative stress · Nrf2 · NF- κ B · HDAC

Sulforaphane

Discovery and isolation

Since 1980s, it was known that the consumption of green and yellow vegetables (leafy green vegetables, cauliflower, carrots, broccoli, Brussel sprouts, etc.) was related to a reduction in the risk of developing certain types of cancer (Colditz et al. 1985). Later, studies showed that some molecules contained in those vegetables were capable to induce xenobiotic metabolism and antioxidant enzyme activation and, as a common feature, those molecules were glutathione S-transferase (GST) substrates, becoming potential protective agents against developing cancer (Talalay et al. 1988). Later, different green and yellow vegetables were tested to determine which of those molecules could activate phase II enzyme activity, with the cruciferous family being the one that enhanced these enzymatic activities the most, particularly broccoli and Brussel sprout extracts (Prochaska et al. 1992). Almost immediately, an isothiocyanate described as “a potent phase II enzymes inducer” was isolated from broccoli extracts, and was identified by spectroscopic methods as 1-isothiocyanato-(4R)-(methylsulfinyl) butane or sulforaphane (SFN); (Zhang et al. 1992). Phase II enzymes are composed of antioxidant and conjugating enzyme which are capable of enhancing xenobiotic hydrophilicity, facilitating their excretion, promoting cell detoxification and

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antioxidant response (Xu et al. 2005). Following the publication of these results, many studies were focused in the SFN synthesis, mechanism of action, and its protective role against tumorigenesis and electrophile attacks provoked by oxidative stress (Prester et al. 1993; Talalay et al. 1995) (Fig. 1).

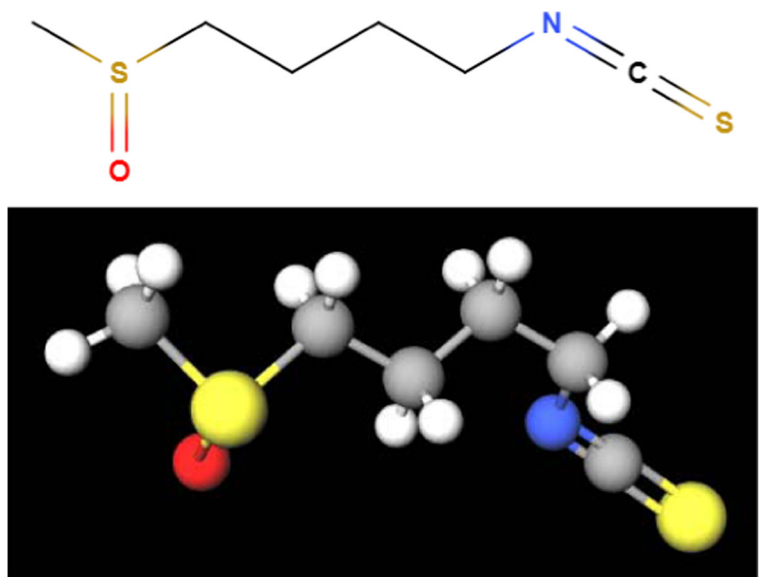
Biosynthesis and other products after glucoraphanin

SFN is the hydrolysis product of glucoraphanin (4-(methylsulfinyl) butyl glucosinolate), one of the main glucosinolates contained in cruciferous vegetables, and the most abundant in broccoli and Brussel sprouts (Ghawi et al. 2013; Brown et al. 2002). Glucosinolates are a family of amino acid-derived secondary metabolites characterized by the presence of three moieties: sulfur groups within a thiohydroximate-*O*-sulfonate structure; a D-glucose molecule; and an α -amino acid-derived alkyl, aralkyl, or indolyl side chain (Barba et al. 2016; Ishida et al. 2014). The glucoraphanin biosynthetic pathway comprises, at least, three stages: (1) a side chain elongation round by deamination of amino acids to a 2-oxo-acid; (2) glucosinolate core structure formation by S-glucosyltransferases; and (3) a S-oxygenation mediated by monooxygenases, to finally get the bioactive form of glucoraphanin as product (Ishida et al. 2014; Yang et al. 2017). The glucoraphanin concentration in the plant mostly depends on the organ, post-germination time, environmental conditions, post-harvest handling conditions, and

storage (Guo et al. 2014; Winkler et al. 2007; Martinez-Villaluenga et al. 2010). When the plant containing glucosinolates is mechanically damaged (cut, chewed, or chopped), or is under stress conditions, such as a bacterial or fungal infection, a hydrolase called myrosinase (thioglycoside glucohydrolase), which is usually separated in a different compartment, is released to interact with glucoraphanin molecules to hydrolyze it, releasing a D-glucose molecule and a thiohydroximate-*O*-sulfate, an unstable aglycone (Burmeister et al. 2000), which spontaneously rearranges to the isothiocyanate form of sulforaphane (Angelino and Jeffery 2014; Barba et al. 2016; Bones and Rossiter 2006) (Fig. 2).

Myrosinase activity decreases with heat exposure when the vegetables are cooked before eating, favoring the presence of glucoraphanin and diminishing the availability of sulforaphane when the vegetables are cooked before consumption (Ghawi et al. 2013). However, myrosinase activity is not only found in plants, it could also be found an equivalent activity in mammal's lower gut microbiota thioglucosidases, allowing the hydrolysis of glucoraphanin and the sulforaphane absorption (Lai et al. 2010; Bheemreddy and Jeffery 2007). Some species of bacteria, such as *Escherichia coli*, *Bacteroides thetaiotaomicron*, *Enterococcus faecalis*, *Enterococcus faecium*, *Peptostreptococcus* sp., and *Bifidobacterium* sp. can process glucosinolates due to the presence of specific thioglucosidases, maintaining the glucoraphanin conversion to isothiocyanates even

Fig. 1 Molecular structure of sulforaphane. The isothiocyanate sulforaphane is an aliphatic small molecule (177.26 g/mol) present in cruciferous vegetables. SFN is shown as a 2D (up) and 3D (down) molecular model. Due to its small molecular weight and its relative high lipophilicity, sulforaphane can easily cross the distinct cellular membranes



after myrosinase heat deactivation (Hullar and Fu 2014) (Fig. 3).

Physicochemical characterization

Sulforaphane is a small molecule with a molecular weight of 177.28 g/mol and its molecular formula is $C_6H_{11}NOS_2$ with a melting point between 58.6 and 91.2 °C (Fishbein and Heilman 2018). Although glucoraphanin is hydrophilic, sulforaphane, like other isothiocyanates, is typically an aliphatic lipophilic molecule (Fahey and Talalay 1999).

Pharmacokinetics

Absorption, distribution, metabolism, and elimination

Due to sulforaphane's small molecular weight and its relatively high lipophilicity, it is rapidly absorbed in the jejunum across the enteric cells after oral administration (Petri et al. 2003). It reaches the highest concentrations in plasma 3 h after consumption

(approximately 0.9 $\mu\text{mol/L}$), and slowly decreases after the second hour, having an approximate half-life of 2.2 h (Hanlon et al. 2008; Hu et al. 2004; Cramer and Jeffery 2011). When SFN is in the cells, it is metabolized by phase II and III enzymes, for example, glutathione S-transferase (GST) to form conjugated products such as sulforaphane-glutathione (SFN-GSH), sulforaphane-cysteine (SFN-Cys), sulforaphane N-acetyl cysteine (SFN-NAC), and sulforaphane-cysteinyl-glycine (SFN-CG), which are thought to be important for the activation of several SFN biological effects (Clarke et al. 2011). The primary sulforaphane metabolism sites are the intestinal walls; the liver, where it is conjugated with GSH; the kidney, where it is conjugated with NAC; and the bladder (Verkerk et al. 2009) and is accumulated mainly in those same organs, and in lower concentrations in plasma, skin, and lung tissues (Bricker et al. 2014). The excretion rate is higher at the 6th hour after administration (Atwell et al. 2015), excreting in urine SFN-NAC as a principal metabolite, and total elimination is reached in the 12th hour post-ingestion (Cramer and Jeffery 2011).

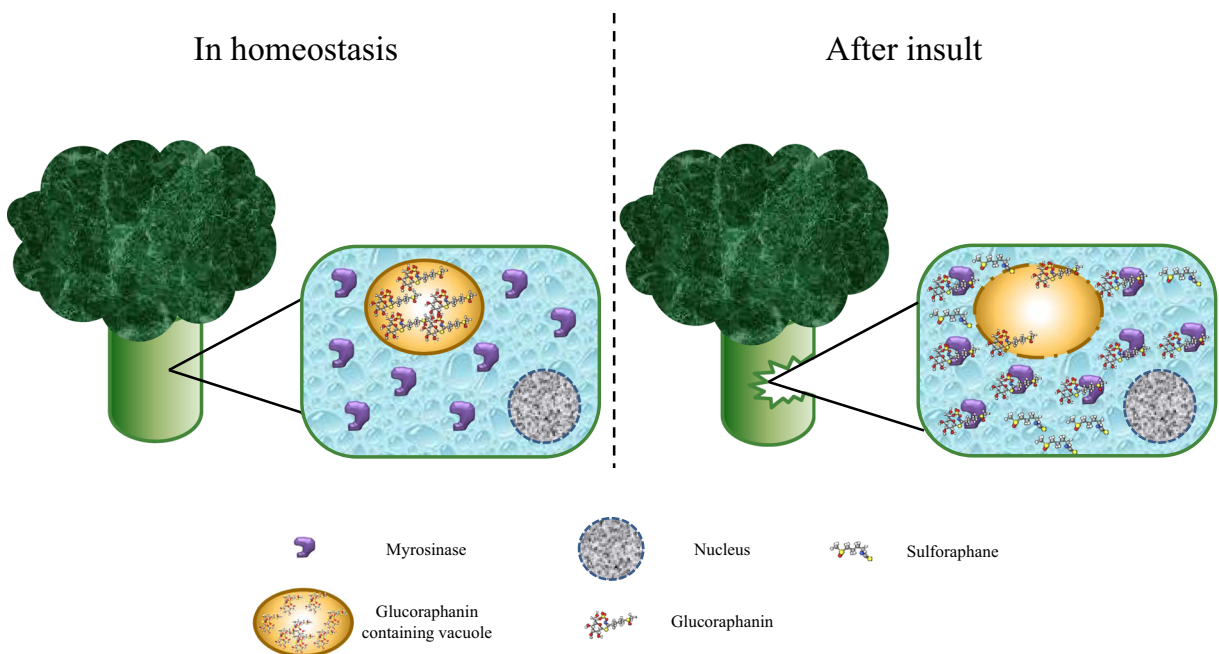


Fig. 2 Sulforaphane synthesis in broccoli. During homeostasis, glucoraphanin, the sulforaphane precursor molecule, is contained in vacuoles inside the broccoli's cells. At the same time, myrosinase, the enzyme responsible of cutting glucoraphanin into glucose and sulforaphane, localizes in the cytosol. Therefore, both

molecules are spatially separated (left panel). When the plant is damaged, i.e., by harvesting, depredation, infection, or even chewing, the vacuoles brake, thus releasing glucoraphanin, which can then interact with myrosinase to form sulforaphane (right panel)

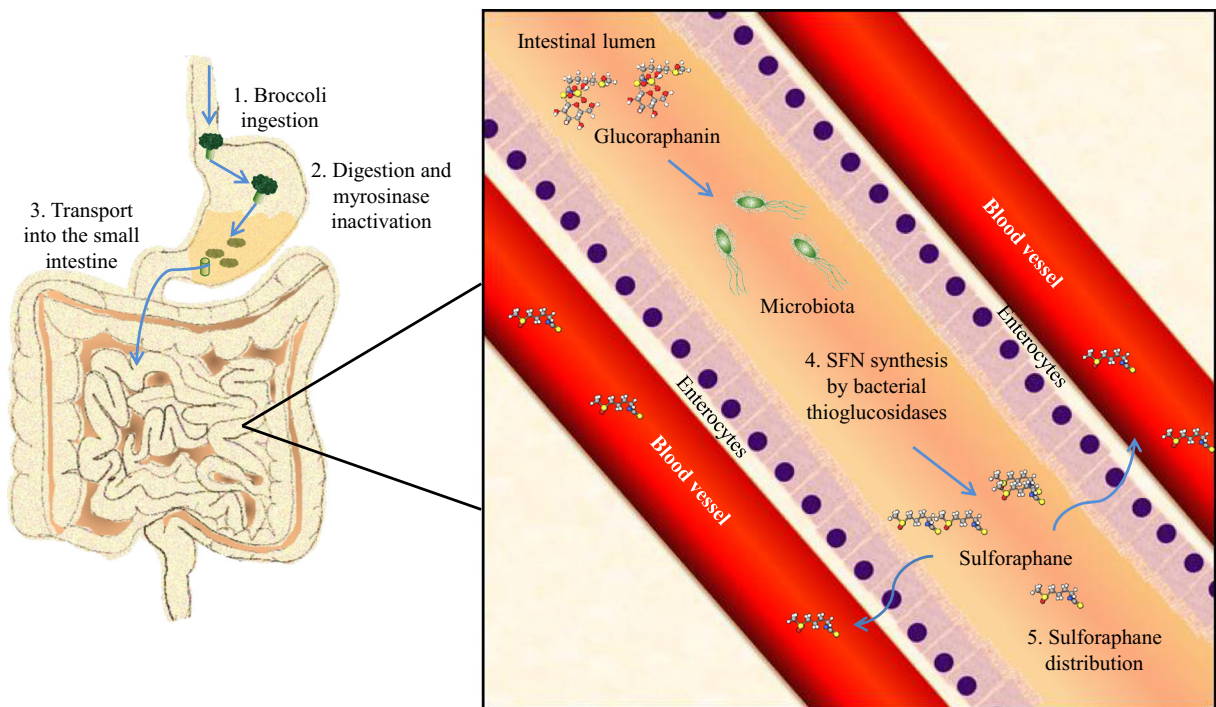


Fig. 3 Sulforaphane synthesis and absorption after consumption. Myrosinase enzymatic activity decreases with heat and low pH, so, it can be denatured when cooked or in the stomach due to the acidic environment, thus diminishing sulforaphane production. When humans consume broccoli, myrosinase activity is usually

abated, conserving glucoraphanin intact until it gets into the small intestine, where specific bacterial species are capable of metabolizing glucoraphanin into sulforaphane through bacterial thioglucosidases, allowing sulforaphane absorption by the enterocytes into the blood flow

Proposed mechanisms of action

Keap1/Nrf2-dependent mechanism

Given that SFN was recognized as a potent inducer of xenobiotic metabolism and antioxidant response, the Keap1/Nrf2 pathway was a very attractive candidate as SFN target. Under basal redox conditions, the nuclear factor erythroid 2-related factor 2 (Nrf2) is sequestered by a Kelch-like ECH-associated protein 1 (Keap1) dimer and rapidly ubiquitinated by the recruitment of the E3 ubiquitin ligase Cul3, resulting in Nrf2 degradation via the proteasome (Li et al. 2004; Silva-Palacios et al. 2018). When the redox state becomes predominantly oxidized, some regulatory cysteines of Keap1 (such as cys151, cys273, and cys288) are oxidized and promote a conformational change that facilitates Nrf2 release and prevents its degradation (Dinkova-Kostova et al. 2017). After being released, Nrf2 requires certain post-translational modifications, namely, tyrosine or serine phosphorylation by

different kinases, such as MAPK or PKC that phosphorylate Nrf2 in Tyr568, as well as the kinase GSK-3 β , which in turn phosphorylates Nrf2 in Ser40. Both phosphorylations are related to Nrf2 nuclear translocation (Bhakkialakshmi et al. 2015). When Nrf2 is imported into the nucleus, it forms a heterodimer with small Maf proteins (MafG, MafK, MafF), which endows it with a DNA-binding capacity to attach to its consensus sequence, the antioxidant response element (ARE; Silva-Palacios et al. 2018), resulting in the transcription of diverse antioxidant response genes, such as NAD(P)H quinone dehydrogenase 1 (NQO1), heme oxygenase 1 (HO-1), and γ -glutamylcysteine ligase (γ GCL), among many others (Guerrero-Beltrán et al. 2012).

In vivo experiments have demonstrated an increased Nrf2 expression and nuclear localization after SFN treatment (Bai et al. 2013), as well as an augmented transcriptional activity (Pu et al. 2018; Zhao et al. 2016). It has also been proposed that SFN is able to oxidize the regulatory cys151 in the Keap1 dimer, thus stopping Nrf2 degradation (Hu et al. 2011).

Interestingly, the Nrf2-dependent antioxidant response is diminished during aging (Zhou et al. 2018), but the SFN treatment has been shown to increase Nrf2 transcription, activation, nuclear translocation, DNA-binding, and antioxidant gene expression in epithelial cells isolated from old rats and elderly humans (Kubo et al. 2017). Therefore, SFN has been proven to be an important inducer of the antioxidant and protective response during aging.

Keap1/Nrf2-independent mechanisms

NF- κ B mechanism

Although SFN administration has been mainly related to the antioxidant response, it has been demonstrated that it is also involved in the regulation of the inflammatory response via the NF- κ B pathway. NF- κ B pathway is considered to be the classical signaling for the inflammatory process and it is present in all cell types. This pathway is involved in the transcription and regulation of different pro- and anti-inflammatory mediators, depending on the activator stimuli. NF- κ B can be activated by two different pathways: the “canonical” and the “alternative,” keeping in common the presence of different inhibitors from the I κ B family (transcriptional repressors that halt NF- κ B in the cytosol), and the I κ B kinase family (IKK; proteins involved in the I κ B phosphorylation and subsequent degradation; Lawrence 2009; Shih et al. 2011; Sun 2017). In mammals, the NF- κ B transcription factor family consists of five distinct proteins: p65 (RelA), RelB, c-Rel, p105/p50 (NF- κ B1), and p100/52 (NF- κ B2), which can form both heterodimers and homodimers, with p50/p65 heterodimer being the most abundant (Oeckinghaus and Ghosh 2009).

Although it is not fully elucidated how SFN inhibits the inflammatory response, it is known that SFN is capable to diminish NF- κ B nuclear translocation and DNA-binding capacity (Heiss et al. 2001). Inflammation master regulators, like TNF- α or IL-6, also decrease after SFN treatment, suggesting an inhibition of the NF- κ B pathway (Negi et al. 2011). Even the secretion of several pro-inflammatory cytokines such as IL-2, IL-4, IL-6, and IFN- γ is inhibited in a SFN dose-dependent manner (Checker et al. 2015), suggesting that NF- κ B could be directly interacting with SFN. However, the effect of this isothiocyanate in the inflammatory response can be observed upon its effect on other proteins

different from NF- κ B, such as I κ B, the NF- κ B repressor responsible for its cytosolic localization, whose degradation is inhibited in a dose-dependent manner in vivo after SFN treatment (Nallasamy et al. 2014) (Fig. 4).

Epigenetic mechanism

Another mechanism of action for SFN was proposed several years ago by Myzak, Ho, and Dashwood. Knowing that SFN-NAC (SFN-N-Acetylcysteine) and SFN-Cys are important metabolites formed when SFN is metabolized through the mercapturic acid pathway, and using a molecular modeling approach, they discovered a plausible interaction for the carboxylate group of SFN-Cys within the active site of HDACs (histone deacetylases) using Zn as a bidentate ligand (Myzak et al. 2004). This interaction of SFN metabolites inhibits the HDAC activity, thus altering the cellular epigenetic pathways.

The same research group established an association between HDAC inhibition and histone acetylation increase on Bax and p21 promoters (Myzak et al. 2006b); Bax is a well-known antiapoptotic protein that belongs to the Bcl₂ family, while p21 is a cell cycle inhibitor that binds to CDK-cyclins and blocks their effect. So, elevated Bax and p21 mRNA and protein expression have been associated with cell cycle arrest and apoptotic cell death in cancer cells after SFN treatment due to an epigenetic mechanism (Myzak et al. 2006a, 2006b) (Fig. 5).

Currently, it is also known that this particular isothiocyanate and its metabolites decrease the expression of DNA methyltransferases (DNMTs), especially DNMT1 and DNMT3b. In particular, SFN diminishes methylation in cyclin D2 promoter regions containing c-Myc and multiple Sp1 binding sites (Hsu et al. 2011). These results, together with SFN HDAC inhibitory effect, place these molecules as important epigenetic regulators, which are able to induce the transcriptional activation of several tumor suppressor genes (Dashwood and Ho 2008).

Being SFN, an isothiocyanate derived from cruciferous vegetables; it has become an important object of investigation as a functional food or as a food supplement. To date, there are numerous studies where SFN improves cellular conditions due to its epigenetic role.

The effect of SFN as an HDAC inhibitor was mainly explored to explain its anti-tumorigenic effect using essentially human embryonic kidney 293 cells, HCT116 human colorectal cancer cells and BPH-1, LnCaP and PC-3 prostate epithelial cells, and also some in vivo murine models

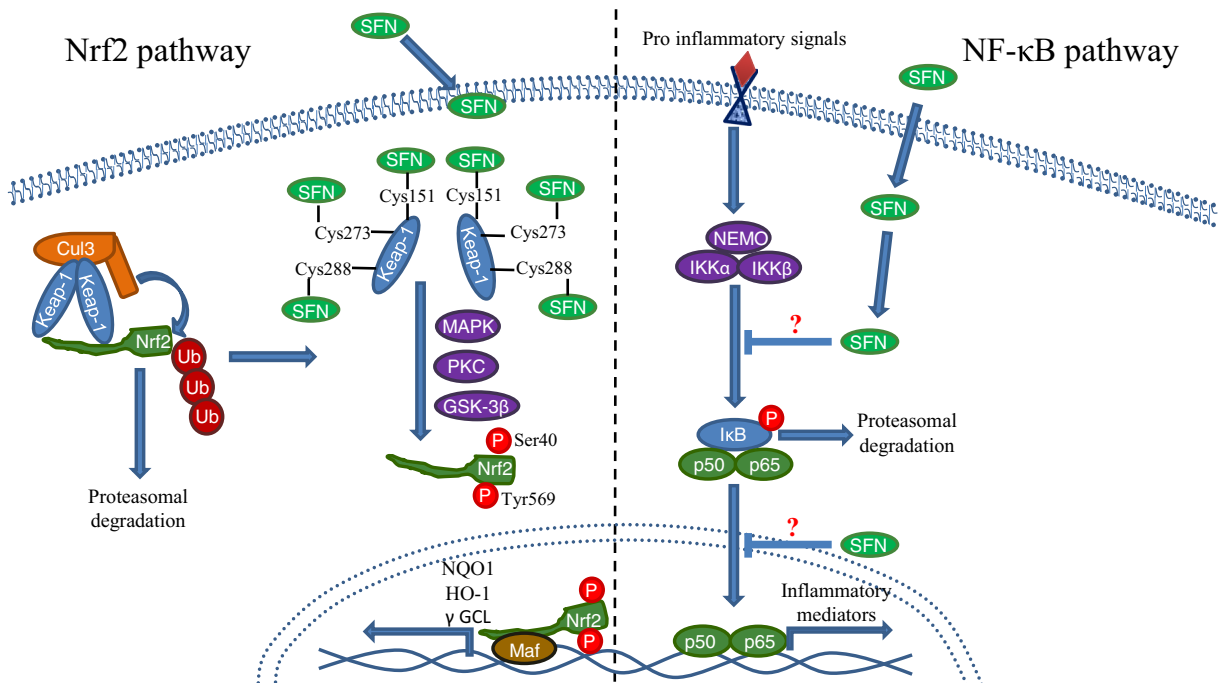


Fig. 4 Effects of sulforaphane on Nrf2 and NF- κ B pathways. After sulforaphane enters the cell, it can interact with many different proteins, directly modulating numerous cellular pathways. Nrf2 pathway: Keap1/Nrf2 pathway is one of the most important pathways affected by sulforaphane that is responsible for antioxidant response and xenobiotic metabolism (left panel). Nrf2 is repressed by Keap1 and degraded by the proteasome under basal conditions, but when Keap1 regulatory cysteines are oxidized, Nrf2 is released and post-translationally modified to be translocated into the nucleus and dimerize with small Maf proteins. Nrf2 can then bind to antioxidant response elements, namely AREs. Also, sulforaphane can oxidize some important regulatory

cysteines from Keap1, allowing Nrf2 cytosolic accumulation and nuclear importing. NF- κ B pathway: NF- κ B is also an important pathway regulated by sulforaphane. It is in charge of inflammatory response regulation. Under basal conditions, NF- κ B is sequestered into the cytosol by I κ B, but when pro-inflammatory ligands bind to its receptors, the IKK protein family phosphorylates I κ B to degrade it via proteasome, so NF- κ B is able to translocate into the nucleus and transcribe several inflammatory mediators. Sulforaphane is capable to inhibit I κ B phosphorylation and NF- κ B nuclear translocation, although the exact mechanism of action is not fully known

(Myzak et al. 2006b), where SFN was shown to increase the acetylation of histone H3 and histone H4.

SFN is also known to inhibit TERT expression by decreasing DNMTs in MCF-7 and MDA-MB-231 human breast cancer cells (Meeran et al. 2010); therefore, SFN alone or in combination with other natural compounds such as genistein (Paul et al. 2018), withaferin A (Royston et al. 2017), or epigallocatechin-3-gallate (Li et al. 2016) inhibits cellular proliferation of breast cancer cell lines.

Even though there are many reports related to SFN's neuroprotective effect in brain cells and neurodegenerative diseases, most of them are related to its effects associated to Nrf2 activation, which were already discussed above. Nevertheless, there are very few studies addressing SFN's epigenetic mechanism in the brain. More specifically, Zhao et al. (2018) found that SFN

augmented the antioxidant and anti-inflammatory capacity of mouse neuroblastoma N2a cells that overexpress the human Swedish mutant amyloid precursor protein (N2a/APP^{swe} cells), which is a common cellular model of Alzheimer's disease. They found that SFN upregulated Nrf2 expression by reducing DNA demethylation levels of the Nrf2 promoter (Zhao et al. 2018), in a very interesting study that involves both the epigenetic mechanism and the Nrf2-activation mechanism. In another model using the triple-transgenic mouse model of Alzheimer's disease (3 \times Tg-AD), Kim et al. (2017) showed that the use of SFN regulates the expression of the Brain-derived neurotrophic factor (BDNF) via HDAC inhibition, thus increasing H3 and H4 acetylation on the BDNF promoter. Enhancing BDNF expression as an effect of SFN treatment increased the neuronal content of several synaptic molecules like MAP 2,

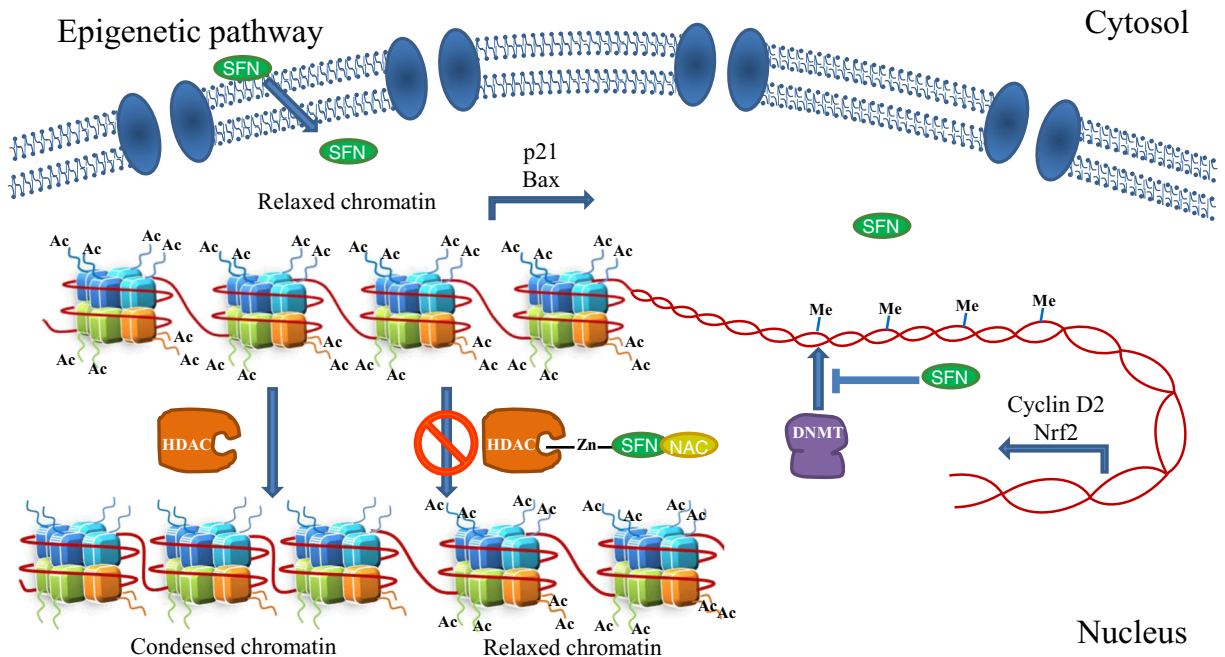


Fig. 5 Effects of sulforaphane on epigenetic mechanisms. Sulforaphane is able to interact with HDACs, modifying the chromatin compaction state. It is known that sulforaphane linked to NAC can bind to the HDAC's active site using a Zn atom, thus inhibiting histone deacetylation. The chromatin relaxation allows

transcription of certain genes such as p21 or Bax, which act as tumor suppressors (left). Sulforaphane can also decrease DNMT expression, thus inducing various gene transcription, such as cell cycle regulators (like Cyclin D2) or stress-protector transcription factors (like Nrf2; right)

synaptophysin, and PSD-95 in primary cortical neurons of $3 \times$ Tg-AD, suggesting that this molecule might be an attractive drug to minimize neurodegenerative disorders. However, more research is needed to evaluate the epigenetic mechanism of SFN in the brain.

Sulforaphane effects on proteasome status

Proteasome in aging and neurodegeneration

Proteasomes are the main cellular degradation machineries implicated in almost all pathways and (normal and pathological) conditions through their role in the maintenance of proteome homeostasis (proteostasis; Papaevgeniou and Chondrogianni 2016). The main eukaryotic proteasome complex is called 20S core that is produced by the association of 4 rings composed by 7 different α - and β -type subunits in an $\alpha_7\beta_7\beta_7\alpha_7$ order. On one or both edges of the 20S core, various regulatory complexes may be bound giving rise to higher molecular weight complexes with the 26S/30S proteasome being the most studied (produced through

binding of one (26S) or two (30S) 19S complexes at the 20S core ends) (Ciechanover 1998; Budenholzer et al. 2017).

Apart of the role of this multi-enzyme in the regulation of almost all vital cellular processes such as the cell cycle and protein catabolism, it is also in charge of the cellular detoxification from oxidized and misfolded proteins that accumulate during the progression of aging or age-related diseases (Chondrogianni et al. 2015). We, among others, have shown that the proteasome function is reduced during the progression of cellular senescence and aging (Chondrogianni et al. 2000, 2003, 2008). Consequently, proteasome activation has emerged as a hot spot in the anti-aging field (Chondrogianni et al. 2015b; Vilchez et al. 2014). More specifically, proteasome activation in primary fibroblasts either through genetic overexpression of proteasome subunits (Chondrogianni et al. 2005; Chondrogianni and Gonos 2007; Hwang et al. 2007) or through natural compound treatment (Katsiki et al. 2007; Kapeta et al. 2010; Kwak et al. 2007) has been shown to lead to increased cellular lifespan and enhanced stress resistance. The same was

Effects on proteasomal function

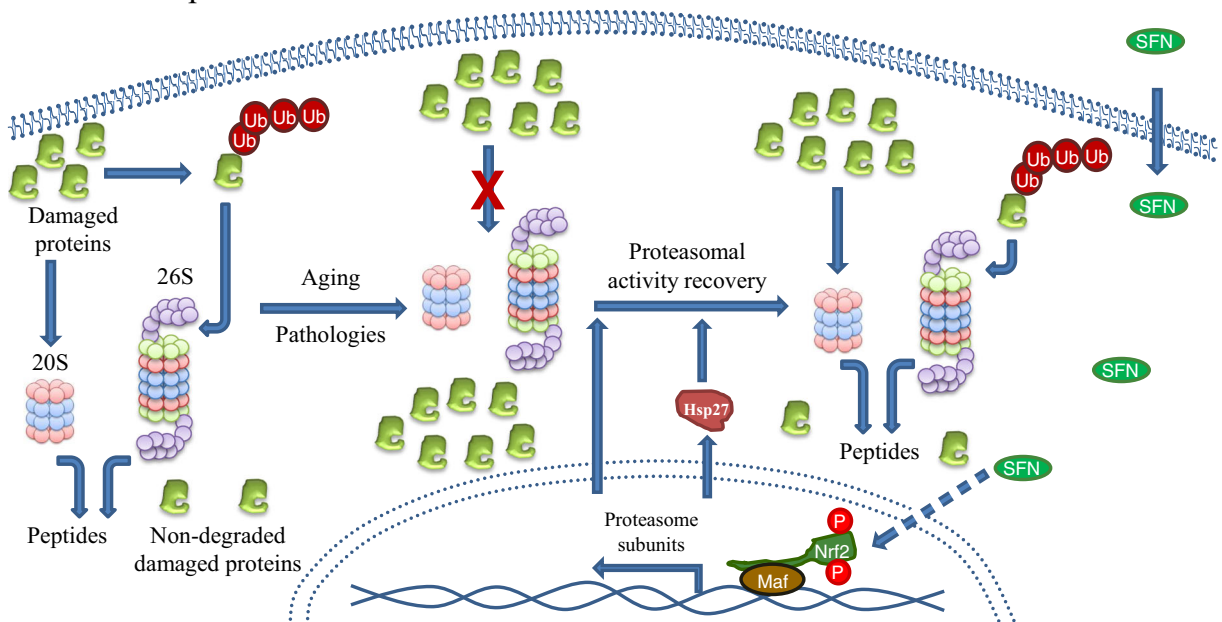


Fig. 6 Effects of sulforaphane on proteasomal activity. Proteasome activity is fundamental for maintaining proteostasis. It is known that impaired proteasome function is related to several pathologies, and occurs naturally during aging as shown in the figure, leading to abnormal damaged protein accumulation and cell stress. Sulforaphane can reactivate proteasome activity and

function during aging and some pathologies through Nrf2 activation. This is due to the Nrf2-mediated transcriptional regulation of many proteasome subunits under stress conditions. At the same time, sulforaphane upregulates HSP70 further enhancing proteasome activation

shown to occur in lens epithelial cells, thus showing that it is not a cell type-specific effect (Liu et al. 2007). More recently, we achieved proteasome activation in the multicellular organismal level (*C. elegans*) through genetic overexpression of *pbs-5* proteasome subunit (Chondrogianni et al. 2015a) as well as through a natural compound that is a diet constituent (Papaevgeniou et al. 2016). On top of the promotion of proteasome activation on the anti-aging field, this strategy is also emerging in the field of aggregation-related diseases (such as Alzheimer's, Parkinson's, Huntington's disease among others) where the proteasome is either inhibited or downregulated upon the disease progression (Vilchez et al. 2014). Overexpression of Rpn6 proteasome subunit in *C. elegans* or Rpn11 proteasome subunit in *D. melanogaster* results in reduced toxicity of poly-Q aggregates and suppressed neurodegeneration in HD models (Vilchez et al. 2012; Park et al. 2009). We have also shown that proteasome activation exerts a protective role against A β toxicity in various AD models of *C. elegans* and in murine primary neurons (Papaevgeniou et al. 2016).

Sulforaphane enhances the proteasomal function

Microarray analysis has shown that most of the proteasome subunits are transcriptional targets of Nrf2 in mammals (Kwak et al. 2003) while the same was also shown for its orthologue namely SKN-1 in *C. elegans* (Kahn et al. 2008). This regulation occurs under specific conditions such as the cellular response to oxidative stress induced by SFN. In addition to the enhanced SFN-mediated induction of several proteasome subunits (Kwak et al. 2007), increased proteasome activity was also attributed to the SFN-mediated upregulation of Hsp27 that was shown to promote proteasome activation as well (Gan et al. 2010) (Fig. 6).

Treatment of murine neuroblastoma cells with SFN enhanced both the expression and the function of proteasome via Nrf2-mediated regulation endowing cells with enhanced resistance against H₂O₂-mediated oxidative stress and toxicity (Kwak et al. 2007). Additionally, SFN-mediated Nrf2 induction protected neuroblastoma cells from A β -induced cell death. This protection was

shown to be dependent on proteasome activation promoted by Nrf2 (Park et al. 2009). Upon oxidative stress adaptation, Nrf2-dependent enhancement of oxidized protein degradation has been attributed to the enhanced expression of both the 20S core and the 11S regulator (Pickering et al. 2012) (Fig. 6).

The proteasome activating effects of SFN were also verified in UPS function reporter mice. On top of the proteasome activation, enhancement of the autophagy activity was also revealed in the brain and the liver of the relative mice, suggesting that this might be another pathway to be explored due to SFN treatment.

Sulforaphane effects on the central nervous system

Sulforaphane in neuroinflammation

Neuroinflammation is a common feature of neurological diseases. Inflammasomes, which form a multiprotein complex within the innate immune system, induce inflammation in response to various stimuli, such as pathogens and stress. Inflammasomes activate pro-inflammatory caspases, like caspase-1, that lead to the activation of pro-inflammatory cytokines, including the interleukins IL-1b, IL-18, and IL-33, which promote neuroinflammation and brain pathologies (Pennisi et al. 2017). The toll-like receptors (TLR4) also participate in the process of neuroinflammation, due to ROS and cytokine generation (Iizumi et al. 2016). Both astroglia and microglia express TLR4 receptors, and endogenous ligands produced in the ischemic brain induce inflammatory responses. It has been reported that hyperammonemia induces neuroinflammation and increases GABAergic secretion. Hernandez-Rabaza et al. (2016) showed that SFN administration to Wistar male rats with hyperammonemia decreased IL-1b and GABA, and increased IL-4 and IL-10. Moreover, it has also been observed that in 2-month-old C57BL/6 mice, in which cognitive functions were decreased due to neuroinflammation induced with LPS for 7 days, pre-treatment with SFN 30 min before LPS administration improved the temporal space behavior and the memory compared to those animals administered only with LPS (Gao et al. 2018).

The most recognized idea to explain the mechanism by which SFN decreases neuroinflammation is the activation of the transcription factor Nrf2 and the increase in

the expression of its target genes, as described previously. For example, in the microglial cell line BV2 and in the primary microglial cultures from adult and aged mice stimulated with LPS to induce an inflammatory state, SFN treatment decreased the expression of pro-inflammatory markers such as IL-1b, IL-6, and iNOS via Nrf2 antioxidant response (Townsend and Johnson 2016). In an okadaic acid model (OKA) (Tonoki et al. 2009), which induces oxidative stress and neuroinflammation by decreasing Nrf2 levels, the administration of SFN prevented the memory impairment induced by OKA in rats. That treatment also restored Nrf2 and antioxidant protein (GCLC, HO-1) expression, diminished the oxidative stress by attenuating ROS and NO levels, and increased GSH concentration. In addition, neuroinflammation was also diminished by reducing NF- κ B and TNF- α , and by rising IL-10 levels, which was followed by a reduction in neuronal apoptosis in the rat's cerebral cortex and hippocampus (Dwivedi et al. 2016).

Conversely, in a dietary intervention experiment, old mice were fed with a 10% broccoli diet for 28 days, and the neuroinflammation was reduced by an increment of interleukin-1 β mRNA and a decrement of b-245 β , preventing the upregulation of reactive glia markers. In this way, a 10% broccoli diet provided a modest reduction in age-related oxidative stress and glial reactivity, but was insufficient to inhibit LPS-induced inflammation (Townsend and Johnson 2016).

Another interesting mechanism proposed to explain SFN participation in the reduction of neuroinflammation is through the inhibition of the transcription factor NF- κ B. In a model of human macrophages (THP-1) exposed to the peptide A β 1-42 to induce an inflammatory state and subsequently treated with SFN, IL-1b and TNF- α levels decreased, and that correlated with a reduction in NF- κ B translocation (Jhang et al. 2018). These results suggest that the effect of SFN might be through a combined mechanism of Nrf2 activation and NF- κ B inhibition in a situation where the cellular redox state could be playing a preponderant role.

Sulforaphane in the treatment of different neuropathologies

Most of the neurodegenerative diseases, including Alzheimer's disease (AD), Parkinson's disease (PD), amyotrophic lateral sclerosis (Bricker et al. 2014), Huntington's disease (Sachdeva et al. 2014), and

multiple sclerosis, are characterized by a compromised redox state, where a deficiency in the antioxidant response, mediated by the transcription factor Nrf2, has generally been observed (Liddell 2017). Since oxidative stress and neuroinflammation have been mainly related to the decrease in cognitive function, it is not surprising that SFN treatment has been used as a conceivable therapy to improved cognitive functions in different animal models.

Several studies have shown that SFN treatment protects against oxidative stress induced by various molecules such as MPTP, 6-OH dopamine, and rotenone in animal models of PD, by increasing Nrf2 activation and regulating the antioxidant response (Jazwa et al. 2011; Morroni et al. 2013; Zhou et al. 2016).

In an HD model using mice treated with 3-NP, SFN treatment decreased the levels of pro-inflammatory cytokines, such as TNF- α , IL-6, iNOS, COX2, as well as NF- κ B activation, and increased the Keap-1/Nrf2/ARE pathway (Jang and Cho 2016). Treatment of cells expressing the mutated huntingtin protein (mHtt) with SFN resulted in increased mHtt degradation that was accompanied by decreased mHtt cytotoxicity. This positive effect was abolished upon treatment with MG132, a proteasome inhibitor, thus further demonstrating the tight interplay between SFN and proteasome activity discussed above (Liu et al. 2014).

Multiple sclerosis (MS) is a chronic disease characterized by focal lesions, axon demyelination, and inflammation that involve the infiltration of peripheral macrophages into the central nervous system. MS is a difficult pathology to study in animal models, so it is usually studied by evaluating the effects of autoimmune responses that resemble MS. The effect of SFN on experimental autoimmune encephalomyelitis (EAE) in C57BL/6 mice significantly inhibited the development and severity of this disease, accompanied by reduced spinal cord inflammatory infiltration and demyelination. In addition, the protection induced by SFN was also related to decrease oxidative stress levels in the mice brains through activation of the Nrf2/ARE pathway and the increase in the HO-1 and NQO1 expression. On the other hand, treatment with SFN inhibited the immune responses mediated by the specific Th17 reactions and augmented the anti-inflammatory response mediated by IL-10 (Li et al. 2013).

In the transgenic PS1V97L mice model of AD, 6-month-old animals treated with SFN for 4 months improved their cognitive function in comparison to aged

match non-treated animals (Hou et al. 2018). In another AD model, the double transgenic amyloid APP/PS1 (precursor protein/presenilin 1) mice, SFN treatment improved the ability of independent exploration in the open field, the environmental adaptability, and ameliorated the Morris maze test (Zhang et al. 2017). Interestingly, consumption of glucoraphanin, a precursor of SFN, by 6-week-old young mice prevented the cognitive function decline during adulthood when phenylcyclidine (an NMDA receptor antagonist) was administered, suggesting that SFN might also be used as a prophylactic treatment to prevent cognitive deficit (Shirai et al. 2015).

More recently, the beneficial effects of SFN on hippocampal-dependent spatial memory impairment induced by post-natal proteasome inhibition was revealed. More specifically, SFN administration was accompanied by Nrf2 nuclear translocation and led to upregulation of β 5 proteasome subunit and thus to a potential enhancement of proteasome function that attenuated the deleterious effects of post-natal proteasome inhibition. Mice pretreated with SFN were rescued of spatial memory impairment during adulthood that was developed in mice treated with the proteasome inhibitor MG132 in the absence of SFN. Additionally, SFN treatment promoted the expression of various molecules that are involved in synaptic plasticity (Sunkaria et al. 2018). Protective effects in the brain against stroke were also shown to occur after SFN preconditioning in the cerebral vasculature, although the proteasome status was not examined under those conditions (Alferi et al. 2013). Nevertheless, the possible therapeutic potential of Nrf2 inducers in vascular disease and aging through the crosstalk of Nrf2 with the proteasome has been suggested (Chapple et al. 2012).

Concerning ALS, there are no conclusive results whether SFN counteracted the dysfunctional effects of the disease; however, it has been observed a slight restoration in Nrf2 levels, which might be an interesting start to treat this pathology (Liddell 2017).

Effects of SFN on aging

Longevity and lifespan

Currently, aging is considered a multifactorial, universal, progressive, and deleterious process that occurs over time in living beings, and that has as a consequence the

decline in the organism's physiological functions, increasing their vulnerability to disease and death (López-Otín et al. 2013; Organization, WHO 2015).

Several studies have reported an increment in oxidative stress and a decay in antioxidant defense mechanisms during aging. This situation decreases the organism's protection against numerous oxidative, chemical, and pathological stresses (Dai et al. 2014; Poprac et al. 2017), since it has been proposed that redox status might function as a mediator between the genome and epigenome (genes expression) and the exposome (environment, diet, life style, etc. (Jones 2015).

One of the features that is known to decrease during aging is the Nrf2 signaling (Sachdeva et al. 2014; Zhang et al. 2015; Zhou et al. 2018). However, some bioactive molecules, like SFN, are able to re-activate the Nrf2 pathway through all the mechanisms discussed above, thus associating SFN treatment with the prevention of aging decline and age-related pathologies (Greco et al. 2011). Nevertheless, the concentrations of SFN used must be taken into consideration, because it is known that SFN has different effects depending on the cell type and organism, an aspect that is still being studied in the aging context.

In a human model of cataracts associated to aging, lens epithelial cells (hLECs) are known to accumulate oxidative damage by decreasing the antioxidant enzymes, particularly a reduction in Prdx6. When those cells were treated with SFN, they increased the Nrf2 activity leading to Prdx6 expression and cytoprotection against UVB-induced injury (Kubo et al. 2017).

Several studies suggest that aging is the result of an age-associated decline in stem cell self-renewal, replication, and lineage commitment (Sharpless and DePinho 2007). When the effects of SFN were evaluated on the biology of human mesenchymal stem cells (MSCs), it was found that low doses of SFN promoted MSC proliferation and protected them from apoptosis and senescence, but higher doses had a cytotoxic and contrary effect. Probably, the adverse effects of SFN were due to its pro-oxidant function generating glutathione depletion and superoxide production (Zanichelli et al. 2012).

It is well known that increased cell senescence contributes to aging-associated diseases. Senescence can be induced by stressors like oxidative stress, dysfunctional telomeres, excessive mitogen signaling, perturbations in chromatin organization, oxidative DNA damage, and oncogenic activation, among others (Campisi and di Fagagna 2007; Childs et al. 2015). Many studies have

shown a continuous increase in glucose metabolism through glycolysis in senescence. Hariton et al. (2018) showed evidence that treatment with 1 μ M SFN once per week delayed the senescence of human MRC-5 and BJ fibroblasts, exhibiting a caloric restriction mimetic-like activity and a decreased oxidative damage to proteins and DNA.

As mentioned above, SFN modulates several epigenetic modifications such as DNA methylation and histone modification (Meeran et al. 2010). It was shown that SFN participates in the inhibition/modulation of HDAC and DNMT activity leading to the reactivation of epigenetically silenced genes in order to enhance chemoprevention (Tortorella et al. 2015). These changes in the epigenome aid in the prevention of neoplasms, resulting in cancer cell death, and potentiate the longevity effects of caloric restriction (Daniel and Tollefsbol 2015).

Regarding premature aging, lately there is an increase interest on studying the protection conferred by SFN against cellular skin aging. Skin aging is sensitive to stress factors including ultraviolet radiation (UVR), excessive alcohol consumption, tobacco abuse, and environmental pollution that lead to cumulative deterioration in skin appearance and function (Pontius and Smith 2011). SFN has demonstrated protective effects against ultraviolet-induced skin damage through several mechanisms of action, by a decrement of oxidative stress (Talalay et al. 2007), and maintenance of collagen levels during photo-aging via the inhibition of the AP-1 activation and the expression of metalloproteinases (Zhu et al. 2004).

Related to the effect of SFN on longevity, in a study of beetles fed with flour diet supplemented with lyophilized broccoli, elongation of their lifespan under physiological conditions (32 °C) as well as during heat-stress conditions (42 °C) was observed. Broccoli also increased longevity in the absence of stress, with the most significant benefit achieved using 1–5% broccoli. The transcription factor Nrf2 increased stress resistance by induction of detoxification, regulation of key stress-resistant factors such as Nrf-2, Jnk-1, and Foxo-1 that resulted to longevity (Grünwald et al. 2013).

The Hutchinson-Gilford progeria syndrome (HGPS) recapitulates the normal aging process, with the expression of mutated lamin A known as 'Progerin' causing DNA damage and genomic instability (Gordon et al. 2014). Progerin is also known to alter various components of the cellular proteolytic pathways, thus leading to impaired proteasome activity and autophagy. Accordingly, the

beneficial effects of SFN on HGPS were also revealed. Fibroblasts from patients and normal individuals were treated with sulforaphane, and progerin clearance was induced (Gabriel et al. 2015). On top of that, the authors showed that long-term treatment of both normal and HGPS fibroblasts with sulforaphane resulted in increased proliferation rates. This was in agreement with the pro-longevity effects of another Nrf2 activator, namely 18 α -glycyrrhetic acid that was shown to act as pro-longevity factor both in normal fibroblasts (Kapeta et al. 2010) and in *C. elegans* (Papaevgeniou et al. 2016).

Conclusions

In summary, SFN has been shown to modulate several cellular pathways in order to activate diverse protective responses, which might allow avoiding cancer and neurodegeneration as well as improving cellular lifespan and health span. However, several interesting questions that still remain unanswered are: Does sulforaphane activate all Nrf2 target genes or is there a selectivity for particular ones such as antioxidant or proteasome genes? If so, how is this selection done? Could SFN be used as an anti-aging compound? Might the Nrf2 induction and NF- κ B repression lead to side effects? What concentrations might be used in cells and animals and for how long? Does SFN have the same effect on young animals as it does on old animals?

Answers to those questions will eventually reveal the therapeutic potential of Nrf2 inducers in general but also the relative protective potential of sulforaphane.

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References

Alfieri A, Srivastava S, Siow RC, Cash D, MODO M, Duchon MR, ... Mann GE (2013) Sulforaphane preconditioning of the Nrf2/HO-1 defense pathway protects the cerebral vasculature

- against blood–brain barrier disruption and neurological deficits in stroke. *Free Radic Biol Med* 65:1012–1022
- Angelino D, Jeffery E (2014) Glucosinolate hydrolysis and bioavailability of resulting isothiocyanates: focus on glucoraphanin. *J Funct Foods* 7:67–76
- Atwell LL, Hsu A, Wong CP, Stevens JF, Bella D, Yu TW, ... Dashwood RH (2015) Absorption and chemopreventive targets of sulforaphane in humans following consumption of broccoli sprouts or a myrosinase-treated broccoli sprout extract. *Mol Nutr Food Res* 59(3):424–433
- Bai Y, Cui W, Xin Y, Miao X, Barati MT, Zhang C et al (2013) Prevention by sulforaphane of diabetic cardiomyopathy is associated with up-regulation of Nrf2 expression and transcription activation. *J Mol Cell Cardiol* 57:82–95
- Barba FJ, Nikmaram N, Roohinejad S, Khelifa A, Zhu Z, Koubaa M (2016) Bioavailability of glucosinolates and their breakdown products: impact of processing. *Front Nutr* 3:24
- Bhakkialakshmi E, Sireesh D, Rajaguru P, Paulmurugan R, Ramkumar KM (2015) The emerging role of redox-sensitive Nrf2–Keap1 pathway in diabetes. *Pharmacol Res* 91:104–114
- Bheemreddy RM, Jeffery EH (2007) The metabolic fate of purified glucoraphanin in F344 rats. *J Agric Food Chem* 55(8):2861–2866
- Bones AM, Rossiter JT (2006) The enzymic and chemically induced decomposition of glucosinolates. *Phytochemistry* 67(11):1053–1067
- Bricker GV, Riedl KM, Ralston RA, Tober KL, Oberyszyn TM, Schwartz SJ (2014) Isothiocyanate metabolism, distribution, and interconversion in mice following consumption of thermally processed broccoli sprouts or purified sulforaphane. *Mol Nutr Food Res* 58(10):1991–2000
- Brown AF, Yousef GG, Jeffery EH, Klein BP, Wallig MA, Kushad MM, Juvik JA (2002) Glucosinolate profiles in broccoli: variation in levels and implications in breeding for cancer chemoprotection. *J Am Soc Hortic Sci* 127(5):807–813
- Budenholzer L, Cheng CL, Li Y, Hochstrasser M (2017) Proteasome structure and assembly. *J Mol Biol* 429(22):3500–3524
- Burmeister WP, Cottaz S, Rollin P, Vasella A, Henrissat B (2000) High resolution X-ray crystallography shows that ascorbate is a cofactor for myrosinase and substitutes for the function of the catalytic base. *J Biol Chem* 275:39385–39393
- Campisi J, di Fagagna F d A (2007) Cellular senescence: when bad things happen to good cells. *Nat Rev Mol Cell Biol* 8(9):729–740
- Chapple SJ, Siow RC, Mann GE (2012) Crosstalk between Nrf2 and the proteasome: therapeutic potential of Nrf2 inducers in vascular disease and aging. *Int J Biochem Cell Biol* 44(8):1315–1320
- Checker R, Gambhir L, Thoh M, Sharma D, Sandur SK (2015) Sulforaphane, a naturally occurring isothiocyanate, exhibits anti-inflammatory effects by targeting GSK3 β /Nrf-2 and NF- κ B pathways in T cells. *J Funct Foods* 19:426–438
- Childs BG, Durik M, Baker DJ, Van Deursen JM (2015) Cellular senescence in aging and age-related disease: from mechanisms to therapy. *Nat Med* 21(12):1424–1435
- Chondrogianni N, Gonos ES (2007) Overexpression of hUMP1/POMP proteasome accessory protein enhances proteasome-mediated antioxidant defence. *Exp Gerontol* 42(9):899–903

- Chondrogianni N, Petropoulos I, Franceschi C, Friguet B, Gonos ES (2000) Fibroblast cultures from healthy centenarians have an active proteasome. *Exp Gerontol* 35(6–7):721–728
- Chondrogianni N, Stratford FL, Trougakos IP, Friguet B, Rivett AJ, Gonos ES (2003) Central role of the proteasome in senescence and survival of human fibroblasts: induction of a senescence-like phenotype upon its inhibition and resistance to stress upon its activation. *J Biol Chem* 278:28026–28037
- Chondrogianni N, Tzavelas C, Pemberton AJ, Nezis IP, Rivett AJ, Gonos ES (2005) Overexpression of proteasome $\beta 5$ subunit increases amount of assembled proteasome and confers ameliorated response to oxidative stress and higher survival rates. *J Biol Chem* 280:11840–11850
- Chondrogianni N, Trougakos IP, Kletsas D, Chen QM, Gonos ES (2008) Partial proteasome inhibition in human fibroblasts triggers accelerated M1 senescence or M2 crisis depending on p53 and Rb status. *Aging Cell* 7(5):717–732
- Chondrogianni N, Voutetakis K, Kapetanou M, Delitsikou V, Papaevgeniou N, Sakellari M, Lefaki M, Filippopoulou K, Gonos ES (2015) Proteasome activation: an innovative promising approach for delaying aging and retarding age-related diseases. *Ageing Res Rev* 23:37–55
- Chondrogianni N, Georgila K, Kourtis N, Tavernarakis N, Gonos ES (2015a) 20S proteasome activation promotes life span extension and resistance to proteotoxicity in *Caenorhabditis elegans*. *FASEB J* 29(2):611–622
- Chondrogianni N, Sakellari M, Lefaki M, Papaevgeniou N, Gonos ES (2015b) Proteasome activation delays aging in vitro and in vivo. *Free Radic Biol Med* 71:303–320
- Ciechanover A (1998) The ubiquitin–proteasome pathway: on protein death and cell life. *EMBO J* 17(24):7151–7160
- Clarke JD, Hsu A, Williams DE, Dashwood RH, Stevens JF, Yamamoto M, Ho E (2011) Metabolism and tissue distribution of sulforaphane in Nrf2 knockout and wild-type mice. *Pharm Res* 28(12):3171–3179
- Colditz GA, Branch LG, Lipnick RJ, Willett WC, Rosner B, Posner BM, Hennekens CH (1985) Increased green and yellow vegetable intake and lowered cancer deaths in an elderly population. *Am J Clin Nutr* 41(1):32–36. <https://doi.org/10.1093/ajcn/41.1.32>
- Cramer JM, Jeffery EH (2011) Sulforaphane absorption and excretion following ingestion of a semi-purified broccoli powder rich in glucoraphanin and broccoli sprouts in healthy men. *Nutr Cancer* 63(2):196–201
- Dai D-F, Chiao YA, Marcinek DJ, Szeto HH, Rabinovitch PS (2014) Mitochondrial oxidative stress in aging and healthspan. *Longev Healthspan* 3(1):6
- Daniel M, Tollefsbol TO (2015) Epigenetic linkage of aging, cancer and nutrition. *J Exp Biol* 218(1):59–70
- Dashwood RH, Ho E (2008) Dietary agents as histone deacetylase inhibitors: sulforaphane and structurally related isothiocyanates. *Nutr Rev* 66:S36–S38
- Dinkova-Kostova AT, Fahey JW, Kostov RV, Kensler TW (2017) KEAP1 and done? Targeting the NRF2 pathway with sulforaphane. *Trends Food Sci Technol* 69:257–269
- Dwivedi S, Rajasekar N, Hanif K, Nath C, Shukla R (2016) Sulforaphane ameliorates okadaic acid-induced memory impairment in rats by activating the Nrf2/HO-1 antioxidant pathway. *Mol Neurobiol* 53(8):5310–5323
- Fahey J, Talalay P (1999) Antioxidant functions of sulforaphane: a potent inducer of phase II detoxication enzymes. *Food Chem Toxicol* 37(9–10):973–979
- Fishbein JC, Heilman JM (2018) *Advances in molecular toxicology*. Elsevier Science
- Gabriel D, Roedel D, Gordon LB, Djabali K (2015) Sulforaphane enhances progerin clearance in Hutchinson–Gilford progeria fibroblasts. *Aging Cell* 14(1):78–91
- Gan N, Wu Y-C, Brunet M, Garrido C, Chung F-L, Dai C, Mi L (2010) Sulforaphane activates heat shock response and enhances proteasome activity through up-regulation of Hsp27. *J Biol Chem* 285(46):35528–35536
- Gao J, Xiong B, Zhang B, Li S, Huang N, Zhan G et al (2018) Sulforaphane alleviates lipopolysaccharide-induced spatial learning and memory dysfunction in mice: the role of BDNF-mTOR signaling pathway. *Neuroscience* 388:357–366
- Ghawi SK, Methven L, Niranjani K (2013) The potential to intensify sulforaphane formation in cooked broccoli (*Brassica oleracea* var. *italica*) using mustard seeds (*Sinapis alba*). *Food Chem* 138(2–3):1734–1741
- Gordon LB, Rothman FG, López-Otín C, Misteli T (2014) Progeria: a paradigm for translational medicine. *Cell* 156(3):400–407
- Greco T, Shafer J, Fiskum G (2011) Sulforaphane inhibits mitochondrial permeability transition and oxidative stress. *Free Radic Biol Med* 51(12):2164–2171
- Grünwald S, Stellzig J, Adam IV, Weber K, Binger S, Boll M, Knorr E, Twyman RM, Vilcinskis A, Wenzel U (2013) Longevity in the red flour beetle *Tribolium castaneum* is enhanced by broccoli and depends on nrf-2, jnk-1 and foxo-1 homologous genes. *Genes Nutr* 8(5):439–448
- Guerrero-Beltrán CE, Calderón-Oliver M, Pedraza-Chaverri J, Chirino YI (2012) Protective effect of sulforaphane against oxidative stress: recent advances. *Exp Toxicol Pathol* 64(5):503–508
- Guo L, Yang R, Wang Z, Guo Q, Gu Z (2014) Glucoraphanin, sulforaphane and myrosinase activity in germinating broccoli sprouts as affected by growth temperature and plant organs. *J Funct Foods* 9:70–77
- Hanlon N, Coldham N, Gielbert A, Kuhnert N, Sauer MJ, King LJ, Ioannides C (2008) Absolute bioavailability and dose-dependent pharmacokinetic behaviour of dietary doses of the chemopreventive isothiocyanate sulforaphane in rat. *Br J Nutr* 99(3):559–564
- Hariton F, Xue M, Rabbani N, Fowler M, Thornalley PJ (2018) Sulforaphane delays fibroblast senescence by curbing cellular glucose uptake, increased glycolysis, and oxidative damage. *Oxidative Med Cell Longev*
- Heiss E, Herhaus C, Klimo K, Bartsch H, Gerhauser C (2001) Nuclear factor- κ B is a molecular target for sulforaphane-mediated anti-inflammatory mechanisms. *J Biol Chem* 276:32008–32015
- Hernandez-Rabaza V, Cabrera-Pastor A, Taoro-Gonzalez L, Gonzalez-Usano A, Agusti A, Balzano T et al (2016) Neuroinflammation increases GABAergic tone and impairs cognitive and motor function in hyperammonemia by increasing GAT-3 membrane expression. Reversal by sulforaphane by promoting M2 polarization of microglia. *J Neuroinflammation* 13(1):83

- Hou T-T, Yang H-Y, Wang W, Wu Q-Q, Tian Y-R, Jia J-P (2018) Sulforaphane inhibits the generation of amyloid- β oligomer and promotes spatial learning and memory in Alzheimer's disease (PS1V97L) transgenic mice. *J Alzheimers Dis* (preprint) 1–11
- Hsu A, Wong CP, Yu Z, Williams DE, Dashwood RH, Ho E (2011) Promoter de-methylation of cyclin D2 by sulforaphane in prostate cancer cells. *Clin Epigenetics* 3(1):3
- Hu R, Hebbar V, Kim B-R, Chen C, Winnik B, Buckley B, Soteropoulos P, Toliás P, Hart RP, Kong A-NT (2004) In vivo pharmacokinetics and regulation of gene expression profiles by isothiocyanate sulforaphane in the rat. *J Pharmacol Exp Ther* 310(1):263–271
- Hu C, Egger AL, Mesecar AD, Van Breemen RB (2011) Modification of Keap1 cysteine residues by sulforaphane. *Chem Res Toxicol* 24(4):515–521
- Hullar MA, Fu BC (2014) Diet, the gut microbiome, and epigenetics. *Cancer J* (Sudbury, Mass.) 20(3):170
- Hwang JS, Hwang JS, Chang I, Kim S (2007) Age-associated decrease in proteasome content and activities in human dermal fibroblasts: restoration of normal level of proteasome subunits reduces aging markers in fibroblasts from elderly persons. *J Gerontol Ser A Biol Med Sci* 62(5):490–499
- Iizumi T, Takahashi S, Mashima K, Minami K, Izawa Y, Abe T, Hishiki T, Suematsu M, Kajimura M, Suzuki N (2016) A possible role of microglia-derived nitric oxide by lipopolysaccharide in activation of astroglial pentose-phosphate pathway via the Keap1/Nrf2 system. *J Neuroinflammation* 13(1):99
- Ishida M, Hara M, Fukino N, Kakizaki T, Morimitsu Y (2014) Glucosinolate metabolism, functionality and breeding for the improvement of Brassicaceae vegetables. *Breed Sci* 64(1):48–59
- Jang M, Cho I-H (2016) Sulforaphane ameliorates 3-nitropropionic acid-induced striatal toxicity by activating the Keap1-Nrf2-ARE pathway and inhibiting the MAPKs and NF- κ B pathways. *Mol Neurobiol* 53(4):2619–2635
- Jazwa A, Rojo AI, Innamorato NG, Hesse M, Fernández-Ruiz J, Cuadrado A (2011) Pharmacological targeting of the transcription factor Nrf2 at the basal ganglia provides disease modifying therapy for experimental parkinsonism. *Antioxid Redox Signal* 14(12):2347–2360
- Jhang KA, Park J-S, Kim H-S, Chong YH (2018) Sulforaphane rescues amyloid- β peptide-mediated decrease in MerTK expression through its anti-inflammatory effect in human THP-1 macrophages. *J Neuroinflammation* 15(1):75
- Jones DP (2015) Redox theory of aging. *Redox Biol* 5:71–79
- Kahn NW, Rea SL, Moyle S, Kell A, Johnson TE (2008) Proteasomal dysfunction activates the transcription factor SKN-1 and produces a selective oxidative-stress response in *Caenorhabditis elegans*. *Biochem J* 409(1):205–213
- Kapeta S, Chondrogianni N, Gonos ES (2010) Nuclear erythroid factor 2 (Nrf2) mediated proteasome activation delays senescence in human fibroblasts. *J Biol Chem* 285(11):8171–8184
- Katsiki M, Chondrogianni N, Chinou I, Rivett AJ, Gonos ES (2007) The olive constituent oleuropein exhibits proteasome stimulatory properties in vitro and confers life span extension of human embryonic fibroblasts. *Rejuvenation Res* 10(2):157–172
- Kim J, Lee S, Choi BR, Yang H, Hwang Y, Park JHY, LaFerla FM, Han JS, Lee KW, Kim J (2017) Sulforaphane epigenetically enhances neuronal BDNF expression and TrkB signaling pathways. *Mol Nutr Food Res* 61(2):1600194
- Kubo E, Chhunchha B, Singh P, Sasaki H, Singh DP (2017) Sulforaphane reactivates cellular antioxidant defense by inducing Nrf2/ARE/Prdx6 activity during aging and oxidative stress. *Sci Rep* 7(1):14130
- Kwak M-K, Wakabayashi N, Greenlaw JL, Yamamoto M, Kensler TW (2003) Antioxidants enhance mammalian proteasome expression through the Keap1-Nrf2 signaling pathway. *Mol Cell Biol* 23(23):8786–8794
- Kwak M-K, Cho J-M, Huang B, Shin S, Kensler TW (2007) Role of increased expression of the proteasome in the protective effects of sulforaphane against hydrogen peroxide-mediated cytotoxicity in murine neuroblastoma cells. *Free Radic Biol Med* 43(5):809–817
- Lai R-H, Miller MJ, Jeffery E (2010) Glucoraphanin hydrolysis by microbiota in the rat cecum results in sulforaphane absorption. *Food Funct* 1(2):161–166
- Lawrence T (2009) The nuclear factor NF- κ B pathway in inflammation. *Cold Spring Harb Perspect Biol* a001651
- Li J, Johnson D, Calkins M, Wright L, Svendsen C, Johnson J (2004) Stabilization of Nrf2 by tBHQ confers protection against oxidative stress-induced cell death in human neural stem cells. *Toxicol Sci* 83(2):313–328
- Li B, Cui W, Liu J, Li R, Liu Q, Xie X-H et al (2013) Sulforaphane ameliorates the development of experimental autoimmune encephalomyelitis by antagonizing oxidative stress and Th17-related inflammation in mice. *Exp Neurol* 250:239–249
- Li Y, Buckhaults P, Cui X, Tollefsbol TO (2016) Combinatorial epigenetic mechanisms and efficacy of early breast cancer inhibition by nutritive botanicals. *Epigenomics* 8(8):1019–1037
- Liddell J (2017) Are astrocytes the predominant cell type for activation of Nrf2 in aging and neurodegeneration? *Antioxidants* 6(3):65
- Liu Y, Liu X, Zhang T, Luna C, Liton PB, Gonzalez P (2007) Cytoprotective effects of proteasome β 5 subunit overexpression in lens epithelial cells. *Mol Vis* 13:31
- Liu Y, Hettinger CL, Zhang D, Rezvani K, Wang X, Wang H (2014) Sulforaphane enhances proteasomal and autophagic activities in mice and is a potential therapeutic reagent for Huntington's disease. *J Neurochem* 129(3):539–547
- López-Otín C, Blasco MA, Partridge L, Serrano M, Kroemer G (2013) The hallmarks of aging. *Cell* 153(6):1194–1217
- Martinez-Villaluenga C, Peñas E, Ciska E, Piskula MK, Kozłowska H, Vidal-Valverde C, Frias J (2010) Time dependence of bioactive compounds and antioxidant capacity during germination of different cultivars of broccoli and radish seeds. *Food Chem* 120(3):710–716
- Meeran SM, Patel SN, Tollefsbol TO (2010) Sulforaphane causes epigenetic repression of hTERT expression in human breast cancer cell lines. *PLoS One* 5(7):e11457
- Morrioni F, Tarozzi A, Sita G, Bolondi C, Moraga JMZ, Cantelli-Forti G, Hrelia P (2013) Neuroprotective effect of sulforaphane in 6-hydroxydopamine-lesioned mouse model of Parkinson's disease. *Neurotoxicology* 36:63–71
- Myzak MC, Karplus PA, Chung F-L, Dashwood RH (2004) A novel mechanism of chemoprotection by sulforaphane: inhibition of histone deacetylase. *Cancer Res* 64(16):5767–5774

- Myzak MC, Dashwood WM, Orner GA, Ho E, Dashwood RH (2006a) Sulforaphane inhibits histone deacetylase in vivo and suppresses tumorigenesis in Apc min mice. *FASEB J* 20(3):506–508
- Myzak MC, Ho E, Dashwood RH (2006b) Dietary agents as histone deacetylase inhibitors. *Mol Carcinog* 45(6):443–446
- Nallasamy P, Si H, Babu PVA, Pan D, Fu Y, Brooke EA et al (2014) Sulforaphane reduces vascular inflammation in mice and prevents TNF- α -induced monocyte adhesion to primary endothelial cells through interfering with the NF- κ B pathway. *J Nutr Biochem* 25(8):824–833
- Negi G, Kumar A, Sharma S, S. (2011) Nrf2 and NF- κ B modulation by sulforaphane counteracts multiple manifestations of diabetic neuropathy in rats and high glucose-induced changes. *Curr Neurovasc Res* 8(4):294–304
- Oeckinghaus A, Ghosh S (2009) The NF- κ B family of transcription factors and its regulation. *Cold Spring Harbor Perspect Biol* a000034
- Organization WH (2015) World report on ageing and health. World Health Organization
- Papaevgeniou N, Chondrogianni N (2016) UPS activation in the battle against aging and aggregation-related diseases: an extended review. In: *Proteostasis*. Springer, pp 1–70
- Papaevgeniou N, Sakellari M, Jha S, Tavemarakis N, Holmberg CI, Gonos ES, Chondrogianni N (2016) 18 α -Glycyrrhetic acid proteasome activator decelerates aging and Alzheimer's disease progression in *Caenorhabditis elegans* and neuronal cultures. *Antioxid Redox Signal* 25(16):855–869
- Park H-M, Kim J-A, Kwak M-K (2009) Protection against amyloid beta cytotoxicity by sulforaphane: role of the proteasome. *Arch Pharm Res* 32(1):109–115
- Paul B, Li Y, Tollefsbol T (2018) The effects of combinatorial genistein and sulforaphane in breast tumor inhibition: role in epigenetic regulation. *Int J Mol Sci* 19(6):1754
- Pennisi M, Crupi R, Di Paola R, Ontario ML, Bella R, Calabrese EJ, ... Calabrese V (2017) Inflammasomes, hormesis, and antioxidants in neuroinflammation: role of NRLP3 in Alzheimer disease. *J Neurosci Res* 95(7):1360–1372
- Petri N, Tannergren C, Holst B, Mellon FA, Bao Y, Plumb GW, ... Knutson L (2003) Absorption/metabolism of sulforaphane and quercetin, and regulation of phase II enzymes, in human jejunum in vivo. *Drug Metab Dispos* 31(6):805–813
- Pickering AM, Linder RA, Zhang H, Forman HJ, Davies KJ (2012) Nrf2-dependent induction of proteasome and Pa28 $\alpha\beta$ regulator are required for adaptation to oxidative stress. *J Biol Chem* 287(13):10021–10031
- Pontius AT, Smith PW (2011) An antiaging and regenerative medicine approach to optimal skin health. *Facial Plast Surg* 27(01):029–034
- Poprac P, Jomova K, Simunkova M, Kollar V, Rhodes CJ, Valko M (2017) Targeting free radicals in oxidative stress-related human diseases. *Trends Pharmacol Sci* 38(7):592–607
- Prestera T, Zhang Y, Spencer SR, Wilczak CA, Talalay P (1993) The electrophile counterattack response: protection against neoplasia and toxicity. *Adv Enzym Regul* 33:281–296
- Prochaska HJ, Santamaria AB, Talalay P (1992) Rapid detection of inducers of enzymes that protect against carcinogens. *Proc Natl Acad Sci* 89(6):2394–2398
- Pu D, Zhao Y, Chen J, Lv A, Zhu S, Luo C, ... Xiao Q (2018) Protective effects of sulforaphane on cognitive impairments and AD-like lesions in diabetic mice are associated with the upregulation of Nrf2 transcription activity. *Neuroscience* 381:35–45
- Royston KJ, Udayakumar N, Lewis K, Tollefsbol TO (2017) A novel combination of withaferin A and sulforaphane inhibits epigenetic machinery, cellular viability and induces apoptosis of breast cancer cells. *Int J Mol Sci* 18(5):1092
- Sachdeva MM, Cano M, Handa JT (2014) Nrf2 signaling is impaired in the aging RPE given an oxidative insult. *Exp Eye Res* 119:111–114
- Sharpless NE, DePinho RA (2007) How stem cells age and why this makes us grow old. *Nat Rev Mol Cell Biol* 8(9):703–713
- Shih VF-S, Tsui R, Caldwell A, Hoffmann A (2011) A single NF κ B system for both canonical and non-canonical signaling. *Cell Res* 21(1):86–102
- Shirai Y, Fujita Y, Hashimoto R, Ohi K, Yamamori H, Yasuda Y, ... Takeda M (2015) Dietary intake of sulforaphane-rich broccoli sprout extracts during juvenile and adolescence can prevent phencyclidine-induced cognitive deficits at adulthood. *PLoS One*. 10(6):e0127244
- Silva-Palacios A, Ostolga-Chavarria M, Zazueta C, Königsberg M (2018) Nrf2: molecular and epigenetic regulation during aging. *Ageing Res Rev* 47:31–40
- Sun S-C (2017) The non-canonical NF- κ B pathway in immunity and inflammation. *Nat Rev Immunol* 17(9):545–558
- Sunkaria A, Bhardwaj S, Yadav A, Halder A, Sandhir R (2018) Sulforaphane attenuates postnatal proteasome inhibition and improves spatial learning in adult mice. *J Nutr Biochem* 51: 69–79
- Talalay P, De Long MJ, Prochaska HJ (1988) Identification of a common chemical signal regulating the induction of enzymes that protect against chemical carcinogenesis. *Proc Natl Acad Sci* 85(21):8261–8265
- Talalay P, Fahey JW, Holtzclaw WD, Prestera T, Zhang Y (1995) Chemoprotection against cancer by phase 2 enzyme induction. *Toxicol Lett* 82:173–179
- Talalay P, Fahey JW, Healy ZR, Wehage SL, Benedict AL, Min C, Dinkova-Kostova AT (2007) Sulforaphane mobilizes cellular defenses that protect skin against damage by UV radiation. *Proc Natl Acad Sci* 104(44):17500–17505
- Tonoki A, Kuranaga E, Tomioka T, Hamazaki J, Murata S, Tanaka K, Miura M (2009) Genetic evidence linking age-dependent attenuation of the 26S proteasome with the aging process. *Mol Cell Biol* 29(4):1095–1106
- Tortorella SM, Royce SG, Licciardi PV, Karagiannis TC (2015) Dietary sulforaphane in cancer chemoprevention: the role of epigenetic regulation and HDAC inhibition. *Antioxid Redox Signal* 22(16):1382–1424
- Townsend BE, Johnson RW (2016) Sulforaphane induces Nrf2 target genes and attenuates inflammatory gene expression in microglia from brain of young adult and aged mice. *Exp Gerontol* 73:42–48
- Verkerk R, Schreiner M, Krumbein A, Ciska E, Holst B, Rowland I, ... Mithen R (2009) Glucosinolates in Brassica vegetables: the influence of the food supply chain on intake, bioavailability and human health. *Mol Nutr Food Res* 53(S2):S219–S219
- Vilchez D, Morante I, Liu Z, Douglas PM, Merkwirth C, Rodrigues AP, ... Dillin A (2012) RPN-6 determines *C. elegans* longevity under proteotoxic stress conditions. *Nature* 489(7415):263

- Vilchez D, Saez I, Dillin A (2014) The role of protein clearance mechanisms in organismal ageing and age-related diseases. *Nat Commun* 5:5659
- Winkler S, Faragher J, Franz P, Imsic M, Jones R (2007) Glucoraphanin and flavonoid levels remain stable during simulated transport and marketing of broccoli (*Brassica oleracea* var. *italica*) heads. *Postharvest Biol Technol* 43(1): 89–94
- Xu C, Li CY-T, Kong A-NT (2005) Induction of phase I, II and III drug metabolism/transport by xenobiotics. *Arch Pharm Res* 28(3):249–268
- Yang H, Liu F, Li Y, Yu B (2017) Reconstructing biosynthetic pathway of the plant-derived cancer chemopreventive-precursor glucoraphanin in *Escherichia coli*. *ACS Synth Biol* 7(1):121–131
- Zanichelli F, Capasso S, Cipollaro M, Pagnotta E, Carteni M, Casale F, Iori R, Galderisi U (2012) Dose-dependent effects of R-sulforaphane isothiocyanate on the biology of human mesenchymal stem cells, at dietary amounts, it promotes cell proliferation and reduces senescence and apoptosis, while at anti-cancer drug doses, it has a cytotoxic effect. *Age* 34(2): 281–293
- Zhang Y, Talalay P, Cho CG, Posner GH (1992) A major inducer of anticarcinogenic protective enzymes from broccoli: isolation and elucidation of structure. *Proc Natl Acad Sci U S A* 89(6):2399–2403
- Zhang H, Davies KJ, Forman HJ (2015) Oxidative stress response and Nrf2 signaling in aging. *Free Radic Biol Med* 88:314–336
- Zhang J, Zhang R, Zhan Z, Li X, Zhou F, Xing A, Jiang C, Chen Y, An L (2017) Beneficial effects of sulforaphane treatment in Alzheimer's disease may be mediated through reduced HDAC1/3 and increased P75NTR expression. *Front Aging Neurosci* 9:121
- Zhao Z, Liao G, Zhou Q, Lv D, Holthfer H, Zou H (2016) Sulforaphane attenuates contrast-induced nephropathy in rats via Nrf2/HO-1 pathway. *Oxidative Med Cell Longev*
- Zhao F, Zhang J, Chang N (2018) Epigenetic modification of Nrf2 by sulforaphane increases the antioxidative and anti-inflammatory capacity in a cellular model of Alzheimer's disease. *Eur J Pharmacol* 824:1–10
- Zhou Q, Chen B, Wang X, Wu L, Yang Y, Cheng X, ... Sun X (2016) Sulforaphane protects against rotenone-induced neurotoxicity in vivo: involvement of the mTOR, Nrf2, and autophagy pathways. *Sci Rep* 6:32206
- Zhou L, Zhang H, Davies KJ, Forman HJ (2018) Aging-related decline in the induction of Nrf2-regulated antioxidant genes in human bronchial epithelial cells. *Redox Biol* 14:35–40
- Zhu M, Zhang Y, Cooper S, Sikorski E, Rohwer J, Bowden GT (2004) Phase II enzyme inducer, sulforaphane, inhibits UVB-induced AP-1 activation in human keratinocytes by a novel mechanism. *Mol Carcinog* 41(3):179–186

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