

A Review of the Sirtuin System, its Clinical Implications, and the Potential Role of Dietary Activators like Resveratrol: Part 2

Gregory S. Kelly, ND

Abstract

The silent information regulator (SIR) genes (sirtuins) comprise a highly conserved family of proteins, with one or more sirtuins present in virtually all species from bacteria to mammals. In mammals seven sirtuin genes – SIRT1 to SIRT7 – have been identified. Emerging from research on the sirtuins is a growing appreciation that they are a very complicated biological response system that influences many other regulator molecules and pathways in complex manners. Part 1 of this article provided an overview of the mammalian sirtuin system, discussed the dietary, lifestyle, and environmental factors that influence sirtuin activity, and summarized research on the importance of vitamin B3 in supporting sirtuin enzyme activity, as well as the role specifically of the amide form of this vitamin – nicotinamide – to inhibit sirtuin enzyme activity. In Part 2 of this review, clinical situations where sirtuins might play a significant role, including longevity, obesity, fatty liver disease, cardiovascular health, neurological disease, and cancer are discussed. Research on the ability of nutritional substances, especially resveratrol, to influence sirtuin expression and function, and hence alter the courses of some clinical situations, is also reviewed. (*Altern Med Rev* 2010;15(4):313-328)

Introduction

The silent information regulator (SIR) genes (sirtuins) comprise a highly conserved family of proteins, with one or more sirtuins present in virtually all species from bacteria to mammals. In mammals seven sirtuin genes – SIRT1 to SIRT7 – have been identified. These seven sirtuin genes code for seven distinct sirtuin enzymes that act as deacetylases or mono-ADP-ribosyltransferases. All sirtuin enzymes are dependent on oxidized nicotinamide adenine dinucleotide (NAD⁺).

As was discussed in Part 1 of this review, sirtuins: (1) are genes that control other genes, (2) respond in an epigenetic manner to a variety of environmental factors, and (3) are hypothesized to

play a particularly important role in an organism's response to certain types of stress and toxicity. Because of this, sirtuins have drawn interest for situations, including lifespan extension, age-related disorders, obesity, heart disease, neurological function, and cancer. This article reviews research on specific clinical situations where sirtuins may potentially play a role. Research on exogenous methods of influencing sirtuins, such as resveratrol, will also be explored.

Anti-aging (Lifespan Extension)

The sirtuin system appears to be involved in mediating the increase in longevity produced by calorie restriction. Limited available evidence also connects increased expression of SIRT1 with increased lifespan and a more gradual aging process, as well as mitigation of symptoms of aging, in some species. As an example, mice that overexpress SIRT1 have an extended lifespan and maintain lower cholesterol, blood glucose, and insulin levels. They also show increased numbers of mitochondria in their neurons.¹ Conversely, the lifespan of mice lacking SIRT1 is reduced under both normal and calorie-restricted conditions.² Interest in sirtuin-mediated longevity and its apparent involvement in ameliorating some age-related changes in physiology and function resulted in the discovery that resveratrol, and possibly other plant compounds, might affect these areas positively.

In vivo studies report mixed results on the lifespan extending effects of resveratrol. It has variously been reported to increase³ or to have no detectable effect⁴ on yeast lifespan. Some studies have reported increased lifespan, subsequent to resveratrol administration, in the nematode worm (*Caenorhabditis elegans*) and fruit flies (*Drosophila melanogaster*). The lifespan-extension response to

Gregory Kelly, ND – Author of the book *Shape Shift*; co-owner of Health Coach; founding partner of Lifestrive; Senior editor, *Alternative Medicine Review*; past instructor at the University of Bridgeport in the College of Naturopathic Medicine; published articles on various aspects of natural medicine and contributed three chapters to the *Textbook of Natural Medicine*, 2nd edition; teaches courses on weight management, the role of stress in health and disease, chronobiology of performance and health, and mind-body medicine. Correspondence address: 7325 1/2 La Jolla Blvd, La Jolla, CA 92037 Email: gregoryskelly@gmail.com website: healthsceneinvestigation.com

resveratrol appeared to be sirtuin-dependent.⁵ Other research has detected no significant effects of resveratrol on lifespan increase in *Drosophila* or *C. elegans*.⁶ While the reason for the mixed findings in yeast, nematode worms, and fruit flies is not completely clear, a study done with the fruit fly species *Anastrepha ludens* suggests that other factors might influence the response to resveratrol. In this study, resveratrol was reported to have a modest effect on lifespan in females but not males. And this effect was only observed in females when diet composition was within a very narrow range of sugar:yeast ratio, suggesting that any longevity benefit resveratrol might have in this species of fruit fly was both gender- and diet-dependent.⁷

Lifespan has been monitored after resveratrol was fed to fish and mammals. Adding resveratrol to the food of the short-lived seasonal fish *Nothobranchius furzeri* (a maximum recorded lifespan of 13 weeks in captivity), starting in early adulthood, produced a dose-dependent increase of median and maximum lifespan, delayed age-related decay in locomotor activity and cognitive performance, and reduction of neurofibrillary degeneration in the brain.⁸ In mice, the effects of resveratrol on lifespan extension might be dependent on diet composition. Resveratrol was reported to extend the lifespan of mice when fed a high-fat diet that resulted in increased calorie consumption;⁹ however, it had no significant effect in extending lifespan in trials when it was given along with a standard-chow diet.^{10,11}

In the study that detected a lifespan extension effect in mice, resveratrol appeared to protect against some of the deleterious physiological effects of a high-fat diet. Compared to a standard-chow diet, a high-fat diet promotes insulin resistance, hyperglycemia, and dyslipidemia. Resveratrol feeding countered these high-fat diet induced changes. Resveratrol feeding also resulted in changes to other metabolic pathways associated with healthy aging, including reduced insulin-like growth factor-1 (IGF-1) levels, increased AMPK and PGC-1alpha activity, increased mitochondrial number, and improved motor function. These responses appear to be mediated by an epigenetic influence of resveratrol. Gene analysis revealed that a high-fat diet significantly modified the expression of 153 pathways; resveratrol opposed the effects of a high-fat diet in 144 of these.⁹

In the mice studies that did not detect lifespan extension, resveratrol still appeared to counter certain age-related changes in gene expression and physiology in a manner closely mimicking the response to calorie restriction. It induced gene expression profiles in multiple tissues, including the heart, skeletal muscle, and brain, that paralleled those induced by long-term calorie restriction.^{10,11} And by old age, resveratrol-fed mice had greater bone density, aortic elasticity, and motor coordination, while also having reduced albuminuria, inflammation, and cataract formation.¹¹

Limited evidence suggests that persimmon oligomeric proanthocyanidins might have lifespan-extending effects. In a mouse model of age-related dysfunction (senescence-accelerated mouse P8), administration of persimmon oligomeric proanthocyanidins extended lifespan. It also increased SIRT1 expression, suggesting that its effects on lifespan might be secondary to its impact on the sirtuin system.¹²

Quercetin has been reported to extend lifespan in *C. elegans*. While quercetin has been reported to impact the sirtuin system (discussed in Part 1), this longevity response does not appear to be dependent on sirtuins, but rather appears to be related to quercetin's influence on the expression of other genes in this species.¹³

Key words: sirtuins, resveratrol, nicotinamide, niacin, vitamin B3, SIR, SIRT, longevity, anti-aging, antiaging, obesity, weight loss, fatty liver, steatosis, cancer, neurological, insulin resistance, metabolic

Table 1. Lifespan Extension and Resveratrol

Organism	Lifespan Extension Findings
Yeast	Mixed findings
<i>Caenorhabditis elegans</i> (nematode worm)	Mixed findings
<i>Drosophila melanogaster</i> (fruit fly)	Mixed findings
<i>Anastrepha ludens</i> (fruit fly)	Modest effect on females that depended on diet composition
<i>Nothobranchius furzeri</i> (short-lived seasonal fish)	Extends lifespan
<i>Mus</i> species (mice)	Extends lifespan when diet is high-fat chow, but no effect on lifespan for standard chow diet

Melatonin might impact sirtuin-mediated aging effects. In senescence-accelerated mice, SIRT1 is significantly lower, as is deacetylation of some of its target proteins. These changes are associated with accelerated aging. Melatonin (10 mg/kg) added to their drinking water, starting from the end of the first month and continued until the end of the ninth month of life, increased SIRT1 and resulted in improved protein deacetylation.¹⁴

Obesity and Metabolic Syndrome

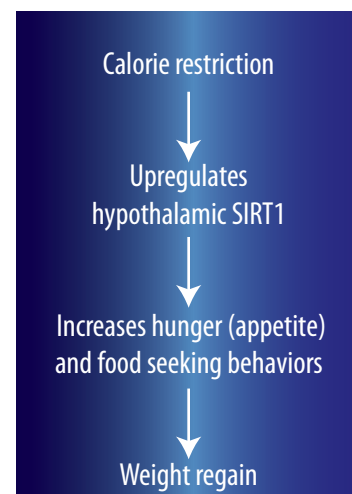
Sirtuins are thought to play a role in obesity and obesity-related issues. Evidence for this role comes from emerging understanding of the regulatory role sirtuins play in metabolic pathways and adaptations linked with obesity and aspects of metabolic syndrome. These include the expression of adipocyte cytokines (adipokines), the maturation of fat cells, insulin secretion and tissue sensitivity, modulation of plasma glucose levels, cholesterol and lipid homeostasis, and mitochondrial energy capacity.¹⁵ SIRT1, for example, is involved in regulating the expression of adipokines such as adiponectin and tumor necrosis factor,¹⁶⁻¹⁸ has been linked to hypothalamic control of energy balance,¹⁹ plays a role in adipogenesis,²⁰ and is involved in the regulation of lipolysis and fatty acid mobilization in response to fasting.²⁰ Evidence from animal experiments where sirtuins are over- or underexpressed, and from limited human evidence, also suggests a role for sirtuins in obesity. Existing evidence on resveratrol suggests that this compound might have sirtuin-mediated anti-obesity effects.

SIRT1 is highly expressed in the hypothalamus (in the arcuate, ventromedial, dorsomedial, and paraventricular nuclei), where it appears to be involved in regulating energy homeostasis, food intake, and body weight.^{19,21} Fasting upregulates hypothalamic SIRT1 expression,²¹ which is associated with the fasting-induced increase in hunger, and is presumably part of the complex adaptations against calorie restriction-induced weight loss. Conversely, pharmacological inhibition of hypothalamic SIRT1 decreases food intake and body weight gain in rodents,¹⁹ suggesting that hypothalamic SIRT1 inhibition might suppress appetite. In mice, calorie restriction induces a complex pattern of physiological and behavioral adaptations, including an increase in activity and food seeking; SIRT1 is required for these behavioral adaptations.²²

In mice, decreased SIRT1 expression in adipose tissue is associated with obesity. In both db/db mice (leptin resistant mice) and mice that have

become obese from eating a high-fat diet, SIRT1 expression in adipose tissue is low.¹⁷ Circumstances that result in SIRT1 underexpression in white adipose enhance adipogenesis and, under fasting conditions, compromise mobilization of fatty acids from white adipocytes. Conversely, circumstances that promote white adipose SIRT1 overexpression are characterized by attenuated adipogenesis and increased lipolysis.²⁰

Figure 1. Hypothalamic SIRT1 Response to Calorie Restriction: Effect on Weight Regain



Experiments with transgenic mice that were bred to moderately overexpress SIRT1 in several tissues also suggest a role for SIRT1 in protecting against obesity. Transgenic mice with greater SIRT1 expression are leaner than littermate controls and have reduced levels of cholesterol, adipokines, insulin, and fasting glucose.^{23,24} Reduced adiposity of these transgenic mice appears to be due to systemic weight regulation that results in decreased whole-body energy requirements, evidenced by the decreased food intake observed in these animals.²³ Although another study did not observe an anti-obesity effect of SIRT1 overexpression in transgenic mice fed a high-fat diet, these mice were protected against some metabolic effects of this diet. Benefits of SIRT1 overexpression included less inflammation, better glucose tolerance, and almost complete protection against hepatic steatosis.²⁵

SIRT1 expression has strong links to insulin sensitivity. Reports indicate that SIRT1 is down-regulated in highly insulin resistant cells, while inducing its expression in these cells increases insulin sensitivity.²³ In skeletal muscle, SIRT1 contributes to the improvement of insulin sensitivity through the transcriptional repression of the protein tyrosine phosphatase 1B (PTP1B) gene.²⁶ In adipocytes, SIRT1 regulates insulin-stimulated glucose uptake and GLUT4 translocation, with greater SIRT1 activity attenuating insulin resistance.¹⁶ In various rodent models of insulin resistance and diabetes, SIRT1 transgenic mice display improved glucose tolerance and insulin sensitivity, due in part to decreased hepatic glucose production and increased hepatic insulin sensitivity.²³ SIRT1 expression appears to improve pancreatic beta-cell function. In beta-cell lines in which SIRT1 expression is inhibited, insulin secretion is blunted. Conversely, increased expression of SIRT1 promotes improved insulin secretion.²⁷ These *in vitro* responses mirror what has been observed *in vivo*. In transgenic mice, bred to overexpress SIRT1 in pancreatic beta-cells, there is enhanced glucose-stimulated insulin secretion and improved glucose tolerance. This improvement of beta-cell function persists through the aging process and when these mice are fed high-fat diets.^{28,29} SIRT1 also regulates cholesterol metabolism by deacetylating and activating LXRalpha, a nuclear receptor involved in cholesterol and lipid homeostasis.³⁰

Less research has been conducted on the other members of the sirtuin family in conditions associated with obesity. The limited evidence suggests that SIRT2 is the most abundant sirtuin in adipocytes, where it appears to be involved in adipogenesis – adipocyte formation. Over expression of SIRT2 inhibits preadipocyte differentiation into adipocytes, while decreased SIRT2 expression promotes adipogenesis.³¹ SIRT3 appears to influence both ATP formation (fatty acid oxidation) and adaptive thermogenesis. In mice lacking SIRT3, fatty acid oxidation disorders emerge during fasting, including reduced ATP levels. These mice also demonstrate a generalized intolerance to cold exposure during fasting, suggesting a disordered thermogenic response from brown adipose.^{32,33} SIRT4 is expressed in beta-cells in the islets of Langerhans and is thought to play a role in mitochondrial regulation of insulin secretion.³⁴ SIRT6 influences the expression of a variety of glycolytic genes, including genes involved in glucose uptake, glycolysis, and mitochondrial respiration. It appears to be a critical element of

glucose homeostasis, with SIRT6-deficient mice developing a lethal hypoglycemia early in life.³⁵ SIRT6 might also play a role in the mouse response to a high-fat diet. Transgenic mice bred to overexpress SIRT6 accumulate significantly less visceral fat and have much lower LDL-cholesterol and triglyceride levels when fed a high-fat diet compared to controls. They also display enhanced glucose tolerance and improved glucose-stimulated insulin secretion.³⁶

In humans, available information on sirtuin interaction with weight has come from observational or calorie restriction studies. In a study of SIRT1 mRNA expression in lean and obese women, lean women were reported to have more than two-fold higher SIRT1 expression in subcutaneous adipose tissue compared to obese women.³⁷ In another study, adipose tissue SIRT1 mRNA expression had a positive association with energy expenditure and insulin sensitivity in 247 nondiabetic offspring of type 2 diabetic patients.³⁸ In a third study, SIRT1-SIRT7 gene and protein expression were determined in peripheral blood mononuclear cells from 54 subjects (41 with normal glucose tolerance and 13 with metabolic syndrome). Insulin resistance and metabolic syndrome were associated with low SIRT1 protein expression.³⁹ In these studies, SIRT1 expression has a negative association with obesity or issues related to obesity; however, whether increased SIRT1 is involved in protecting against obesity, is a marker for obesity resistance, or is altered in response to ongoing dietary, lifestyle, or environmental factors, has not been established and cannot be determined from the existing evidence.

What human evidence does make clear is that, similar to other species including other mammals, human sirtuin expression is sensitive to changes in calorie intake. SIRT1 mRNA was measured in adipose tissue biopsies from nine human volunteers before and after six days of total fasting. Levels in subcutaneous adipose tissue increased more than two-fold with fasting.³⁷ In another study, muscle biopsies were obtained at baseline and on day 21 from 11 nonobese men and women who underwent three weeks of alternate day fasting; a statistically significant increase in muscle SIRT1 mRNA expression was observed.⁴⁰ In a third study, diet-induced changes in adipose tissue gene expression were assessed in two sets of 47 obese women who were placed on either a low-fat (high-carbohydrate) or a moderate-fat (low-carbohydrate) hypoenergetic diet for 10 weeks. One thousand genes, including sirtuin genes, were

regulated by energy restriction. SIRT3 gene expression appeared to be sensitive to the fat-to-carbohydrate ratio of a restricted calorie diet, with increased expression during the moderate-fat diet.⁴¹

Resveratrol has been shown to have *in vitro* and *in vivo* effects on sirtuins that are suggestive of a potential anti-obesity effect. One of these is an ability to counteract circumstances, including high glucose or long-chain fatty acid concentrations, that otherwise reduce the expression of SIRT1.³⁹ Resveratrol also inhibits preadipocyte proliferation and differentiation,⁴² decreases lipid accumulation in, and nonesterified fatty acid release from, adipocytes;⁴³ attenuates fat deposition in hepatic cells;⁴⁴ promotes differentiation of mesenchymal stem cells into osteoblasts at the expense of adipocyte formation;⁴⁵ enhances the lipolytic effect of epinephrine in adipose tissue;³⁷ stimulates glucose uptake by skeletal muscle cells;⁴⁶ enhances insulin sensitivity;²⁶ and protects isolated pancreatic islet cells against cytokine-induced cytotoxicity, which allows these cells to maintain normal insulin-secreting responses to glucose.⁴⁷ As previously mentioned, feeding mice resveratrol appears to counter some of the effects of a high-fat diet; protecting against insulin resistance, hyperglycemia, and dyslipidemia.⁹ Another mice study reported similar benefits when resveratrol was added to a high-fat diet for 13 weeks. In addition to improving insulin sensitivity and glucose tolerance, resveratrol-fed mice had increased metabolic rate, better physical endurance, and reduced fat mass. Although the study did not attempt to monitor changes in sirtuins, resveratrol did change the activity of other proteins, some of which are known to be deacetylated by the sirtuin system.⁴⁸

Fatty Liver Disease

The sirtuin system has a variety of links to alcoholic and nonalcoholic hepatic steatosis. In general, SIRT1 expression has a negative association with fatty infiltration of the liver in both rodents and humans. In rodents, these associations exist for nonalcoholic and alcoholic hepatic steatosis and appear to be related to inflammation and sirtuin interactions with liver fatty acid oxidation and transport.⁴⁹ Sirtuin-steatosis interactions appear to be mediated, at least in part, by sirtuin deacetylation of other proteins, which subsequently modulates the activity of these proteins and their metabolic targets. For example,

in a cell model of hepatic fatty infiltration, SIRT1 protects against hepatic fat deposition via induction of FOXO1 expression and repression of SREBP1 expression.⁴⁴ It has also been proposed that sirtuin effects on the PPARalpha/PGC-1alpha signaling axis might be involved in the protective association.⁴⁹

In rodents, a high-fat diet plays a significant role in interactions with SIRT1 and nonalcoholic hepatic steatosis. Reduced expression of hepatic SIRT1 proteins appears to predispose mice to high-fat diet induced hepatic steatosis, while increased expression appears to protect against steatosis; this has been demonstrated in several studies. When mice, bred to have reduced expression of hepatic SIRT1, were fed a low-fat diet (5% fat), they were no more likely to have manifestations of fatty liver disease than normal mice. However, as dietary fat levels were increased in the mice with reduced hepatic SIRT1 expression, there was a corresponding increase in hepatic steatosis, with higher levels of dietary fat intake causing worse steatosis. These mice, in addition to significant increase in hepatic steatosis, experienced increased liver inflammation and hepatic lipogenesis, with a reduction in fat export.⁵⁰

As mentioned in Part 1 of this review, sirtuins are both a regulating and a regulated protein. Deleted in breast cancer-1 (DBC1) is one protein with an established ability to regulate SIRT1. Mice bred to have a genetic deletion of DBC1 express increased SIRT1 activity in several tissues, including the liver. When these mice are fed a high-fat diet, they become obese but do not develop the hepatic steatosis and inflammation typically caused by this diet and that generally accompanies diet-induced obesity.⁵¹ While increased SIRT1 expression appears to have a protective role against diet-induced hepatic steatosis, evidence also suggests that a high-fat diet can reduce SIRT1 expression. This suggests that an inability to counter the high-fat diet-induced downregulation of SIRT1 might play a role in susceptibility to diet-induced hepatic steatosis.⁵²

SIRT1 expression might also play a role in fatty liver caused by other factors. Monosodium glutamate (MSG) is used to induce obesity and insulin resistance in mice and also results in increased hepatic steatosis. Coadministration of a pharmacological activator of SIRT1 with MSG administration from ages 6-16 weeks protects against hepatic steatosis in MSG-treated mice, despite having no protective effect on weight gain.⁵³

Resveratrol appears to have a protective effect against hepatic fat infiltration. Wang et al also reported an ability of resveratrol to attenuate fat deposition in hepatic cells, secondary to inhibition of SREBP1 expression.⁴⁴ Hou et al observed a resveratrol-induced increase in SIRT1 deacetylase activity. They also detected effects on AMPK and several of its downstream targets, including acetyl-CoA carboxylase and fatty acid synthase. The net result of resveratrol treatment was prevention of hepatic lipid production – effects that were largely abolished by pharmacological and genetic inhibition of SIRT1 deacetylase activity. These findings suggest that resveratrol protects against fatty infiltration by activating SIRT1, which subsequently influences activity of other proteins and a variety of processes involved in the hepatic regulation of lipids.⁵⁴

There are conflicting reports on the effects alcohol has on hepatic SIRT1. Chronic alcohol administration has been variously reported to decrease^{55,56} and increase SIRT1.⁵⁷ The reason for this conflict is not completely clear, although it might be secondary to diet or other factors that influence SIRT1 expression. For example, Lieber et al reported that alcohol reduced hepatic SIRT1 when the fat in the diet consisted of long-chain triglycerides (LCT); however, replacement of LCT with medium chain triglycerides (MCT) restored hepatic SIRT1 almost to levels found without alcohol.⁵⁶ You et al reported that a high saturated-fat diet (40% of energy from cocoa butter) protected against the development of alcoholic fatty liver in mice, while a high polyunsaturated-fat diet (40% of energy from corn oil) did not. The protective effect appeared to be related to sirtuins because, compared with control mice, a diet high in saturated fat upregulated SIRT1 expression and suppressed the ethanol-induced increase in SREBP1, while the corn oil diet did not.⁵⁸ Despite the inconsistent response, available rodent research suggests that normalizing SIRT1 – increasing it when reduced by alcohol and decreasing it when increased by alcohol – might improve resistance to alcohol-induced fatty liver. This has been demonstrated with resveratrol administration.

Amjo et al reported that SIRT1 activity was inhibited by ethanol. Resveratrol treatment increased SIRT1 expression in the liver of ethanol-fed mice. This increase was associated with suppression of SREBP1 and activation of PGC-1 α . Resveratrol also reduced lipid synthesis, increased

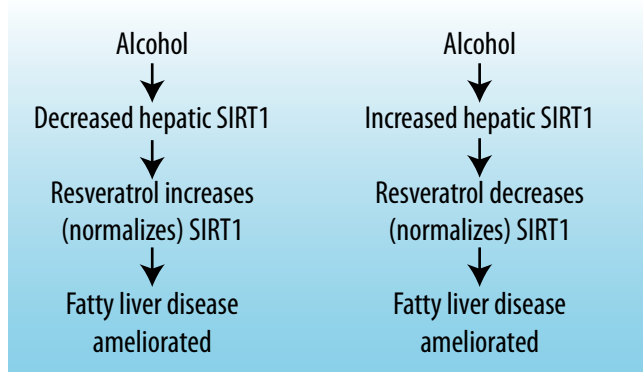
rates of fatty acid oxidation, and prevented alcoholic liver steatosis.⁵⁵ You et al reported that chronic ethanol feeding downregulated hepatic SIRT1 in mice. The reduced expression of SIRT1, since it was unavailable to deacetylate SREBP1, caused an upregulation of this protein. Treatment with resveratrol countered alcohol-induced effects on these regulatory proteins and protected against alcohol-induced fatty liver.⁵⁹ Oliva et al reported an opposite SIRT1 response to alcohol, but still observed a normalizing effect of resveratrol. One month of intragastric feeding of alcohol increased SIRT1 and led to steatosis. Treating alcohol-fed rats with resveratrol inhibited hepatic increase in SIRT1 and, while it was unable to prevent alcohol-induced macrovesicular steatosis, it did protect against necrosis and fibrosis. Hepatic SIRT3 expression was also upregulated by ethanol; resveratrol countered this increase.⁵⁷ These studies implicate sirtuins in alcohol-induced fatty liver disease, and suggest that resveratrol has the potential to help normalize hepatic SIRT1 and other proteins and protect against alcohol-induced fatty liver.

While studies report a mixed response of hepatic SIRT1 to alcohol, resveratrol administration appears to exert an adaptogenic effect by normalizing this response whether alcohol induced an increase or decrease of hepatic SIRT1.

Evidence of interactions with other members of the sirtuin family and fatty liver is sparse. *In vitro*, the number of lipid droplets in human hepatic cells overexpressing SIRT3 was significantly lower than that in control cells. Decreasing SIRT3 expression promoted lipid accumulation in these cells.⁶⁰ Under *in vivo* fasting conditions, SIRT3 expression prevents the accumulation of lipid droplets in hepatic cells.^{32,60,61} Chronic alcohol-feeding also reduced SIRT5.⁵⁶

In humans, SIRT1 expression in visceral adipose tissue was associated with severity of hepatic steatosis. In this study, morbidly obese individuals were divided into two groups – one with moderate hepatic steatosis and the other with severe steatosis. When comparing the two groups, a decrease of SIRT1 mRNA in visceral adipose tissue was detected in samples taken from the group with severe hepatic steatosis. Statistical analysis also revealed a positive correlation between mRNA expression of SIRT1 and homeostasis model assessment for insulin resistance (HOMA-IR).⁶² The researchers did not explore whether the downregulation of SIRT1 mRNA expression in

Figure 2. Alcohol, Sirtuins, Resveratrol, and Fatty Liver



While studies have reported mixed response of hepatic SIRT1 to alcohol, resveratrol administration appears to normalize this response, whether alcohol leads to an increase or decrease of hepatic SIRT1. This suggests adaptogenic activity of resveratrol.

visceral adipose tissue was promoting steatosis in these obese individuals or a response to severe steatosis.

Cardiovascular System

In vitro and *in vivo* evidence suggests a role for several of the sirtuins in the cardiovascular system. SIRT1 appears to play a regulatory role in endothelial function. It is highly expressed in the vasculature, especially during periods of active blood vessel growth and vascular remodeling, when it appears to be involved in angiogenic activity of endothelial cells.^{63,64} SIRT1 promotes endothelium-dependent vasodilation and regenerative functions in endothelial and smooth muscle cells of the vascular wall by targeting endothelial nitric oxide synthase for deacetylation, which stimulates the activity of this enzyme and increases endothelial nitric oxide production. If SIRT1 deacetylation is inhibited in endothelial tissue, nitric oxide synthase acetylation predominates, nitric oxide production decreases, and vasodilation is impaired.⁶⁵ SIRT1 might also play a significant role on endothelial function when blood sugar is elevated. Treatment of human endothelial cells with glucose decreases SIRT1 expression, induces endothelial dysfunction, and accelerates endothelial senescence. Increasing SIRT1 activity inhibits this glucose-induced endothelial senescence and dysfunction. These effects were also seen *in vivo*; activation of SIRT1 prevented hyperglycemia-induced vascular cell senescence and protected

against vascular dysfunction in diabetic mice.⁶⁶

In vitro research suggests resveratrol might augment endothelial SIRT1 expression under circumstances characterized by increased oxidative stress. Exposure of endothelial cells to cigarette smoke extract or hydrogen peroxide decreases SIRT1 levels and enzyme activity with a concomitant increase in acetylated (inactive) nitric oxide synthase. Pretreatment of endothelial cells with resveratrol attenuated the decline in SIRT1 levels and activity and resulted in less acetylation of nitric oxide synthase.⁶⁷ Other research reports resveratrol's endothelial vasoprotective effects⁶⁸ and its decrease in expression of angiotensin II type I receptor in vascular smooth muscle cells *in vivo*. This effect on angiotensin II type I receptors, apparently due to resveratrol's ability to increase expression of SIRT1, blunted angiotensin II-induced hypertension.⁶⁹

SIRT1 might play a role in countering atherosclerosis due to its reported regulation of tissue metalloproteinase 3 (TIMP3). TIMP3 is an endogenous enzyme that counters vascular inflammation and is involved in the prevention of atherosclerosis. SIRT1 activity is also reportedly decreased in atherosclerotic plaques of subjects with type 2 diabetes – a decrease associated with reduced TIMP3 expression.⁷⁰

SIRT1, SIRT3, and SIRT7 are expressed in cardiomyocytes, are upregulated during stress conditions (presumably as an adaptation to counter the stress), and appear to play a critical role in promoting cardiomyocyte resistance to stress and toxicity.⁷¹⁻⁷³ Cardiomyocyte protection appears to occur because of sirtuin deacetylation of other proteins, with the relative balance between acetylation and deacetylation of these targeted proteins influencing whether cardiomyocytes survive under stressful conditions.⁷² Sirtuins also protect cardiomyocytes by activating antioxidant-encoding genes (including manganese superoxide dismutase and catalase) that decrease cellular levels of reactive oxygen species.⁷⁴

Circumstances that result in decreased cardiac SIRT1 are associated with reduced cardiac function. For example, in mice with chronic type 1 diabetes, the enzymatic activity of cardiac SIRT1 is reduced, which contributes to reduced cardiac function and diabetic cardiomyopathy. Resveratrol increases SIRT1 activity and improves cardiac function in these mice.⁷⁵ *In vitro*, resveratrol increases SIRT1 and protects rat cardiomyocytes against hypoxia; pharmacological inhibition of SIRT1 reverses this protection.⁷⁶

Doxorubicin is cardiotoxic, in part because it induces a rapid increase in reactive oxygen species. Pretreatment of cardiomyocytes with resveratrol inhibits the increase in oxidative stress caused by doxorubicin and prevents doxorubicin-induced cardiomyocyte death. These protective effects of resveratrol appear to be sirtuin-mediated, since they are abolished by nicotinamide, an *in vitro* sirtuin inhibitor.⁷⁷

Streptozotocin injections in mice fed a standard-chow diet cause progressive decline in cardiac function associated with markedly reduced cardiomyocyte SIRT1 levels. Adding resveratrol to the diet of these mice increased SIRT1 activity in cardiomyocytes and improved cardiac function.⁷⁵

In rats fed white wine, red wine, resveratrol, hydroxytyrosol, and tyrosol, heart expression of SIRT1 increased to the highest degree with white wine, followed by resveratrol, then tyrosol, hydroxytyrosol, and finally red wine. This was in contrast to the capacity of these dietary additions to offer cardioprotection (gauged by reduction of infarct size and cardiomyocyte apoptosis). Resveratrol provided the most protection, followed in descending order by red wine, hydroxytyrosol, white wine, and tyrosol.⁷⁸

In vitro, nuclear but not cytoplasmic SIRT1 induced the antioxidant enzyme manganese superoxide dismutase, which was further enhanced by resveratrol. Resveratrol's enhancement of enzyme levels suppressed cell death induced by antimycin A or angiotensin II and was dependent on the level of nuclear SIRT1. Oral administration of resveratrol to hamsters also increased manganese superoxide dismutase levels in cardiomyocytes, which then suppressed fibrosis, preserved cardiac function, and significantly improved survival.⁷³

Evidence suggests that resveratrol might help protect against myosin-induced autoimmune myocarditis of rats (a model of human dilated cardiomyopathy). Myosin-immunized rats experience an increase in SIRT1 in the myocardium and in infiltrating mononuclear cells compared with unimmunized rats. Despite the upregulation in SIRT1, myosin-immunization resulted in an increase in heart weight, fibrosis, and the expression of inflammatory cytokines. Resveratrol preserved cardiac function in these rats and protected against cardiomyopathy by decreasing fibrosis and inflammation, while normalizing expression of oxidative stress genes.⁷⁹

While increased cardiomyocyte SIRT1 expression and activity appear to be an adaptation to

stress and toxicity, limited evidence suggests that extremes of increased expression might not be desirable. Transgenic mice bred to have 2.5- to 7.5-fold heart-specific SIRT1 overexpression were protected against oxidative stress. Age-dependent increases in cardiac hypertrophy, apoptosis/fibrosis, cardiac dysfunction, and expression of senescence markers were consequently attenuated. However, a 12.5-fold overexpression of heart-specific SIRT1 increased oxidative stress, apoptosis, and hypertrophy, and decreased cardiac function, stimulating the development of cardiomyopathy.⁸⁰ In this case, rather than being protective and conferring resistance to age-related problems, the highest levels of SIRT1 expression promoted pathology. This may be a result of higher SIRT1 consumption of cellular NAD⁺ exceeding the supply or unbalancing acetylation/deacetylation activities. Whatever the mechanism, these results suggest that the cardioprotective effects of heart-specific SIRT1 expression might be biphasic, with too much expression resulting in diminishing returns (Figure 3).

The importance of other sirtuins for cardiac function is apparent in SIRT3-deficient mice. In these mice, basal levels of ATP in the heart, kidney, and liver are reduced by more than 50 percent, and mitochondrial protein acetylation is markedly elevated in these same tissues. These mice also show signs of cardiac hypertrophy and interstitial fibrosis at age eight weeks and develop severe cardiac hypertrophy in response to hypertrophic stimuli.⁸¹ Conversely, transgenic mice that overexpress SIRT3 are protected from stimuli-induced cardiac hypertrophy.⁷⁴

SIRT7 also appears to be critical for cardiac function. SIRT7-deficient mice have reduced mean and maximum lifespans. Their hearts are characterized by extensive fibrosis, diminished resistance to oxidative and genotoxic stress, and a high basal rate of apoptosis resulting in cardiac hypertrophy and inflammatory cardiomyopathy.⁸²

Brain and Nervous System

Several sirtuins expressed in the mammalian brain appear to play very different roles and respond in dissimilar ways to stress and toxicity. For example, Pfister et al reported that SIRT1 protects neurons against apoptosis, while SIRT2, SIRT3, and SIRT6 induce apoptosis in otherwise healthy neurons. SIRT5 has a dual role. In neurons, where it is located in both the nucleus and cytoplasm, it exerts a protective effect; however, in a subset of neurons where it is located in the mitochondria, it promotes neuronal death.⁸³ While

all these sirtuins appear to impact neurons, almost all research has focused on SIRT1 or SIRT2.

SIRT1 is ubiquitously present in areas of the brain that are especially susceptible to age-related neurodegenerative states (e.g., the prefrontal cortex, hippocampus, and basal ganglia). SIRT1 is also broadly distributed in the neurons that are most susceptible to senescence injury.⁸⁴ Calorie restriction results in upregulation of SIRT1 in some regions of the brain (such as the hypothalamus) and downregulation in others.^{85,86} In mice undergoing calorie restriction, there is an attenuation of beta-amyloid content in the aging brain. This effect can be reproduced in mouse neurons *in vitro* by manipulating cellular SIRT1 expression/activity, suggesting it is a SIRT1-dependent process⁸⁷ and that SIRT1 upregulation might be protective under some types of nutritional stress. SIRT1 is upregulated in primary neurons challenged with some types of neurotoxic insults. However, in transgenic mice created to overexpress human SIRT1 in neurons, the neuronal overexpression of SIRT1 had no neuroprotective effects against damage induced by ischemia or 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine.⁸⁸

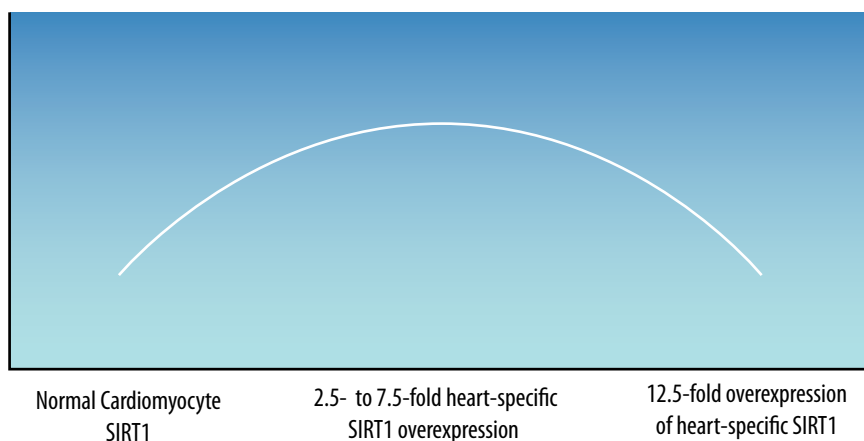
Evidence suggests that SIRT1 is upregulated in the brain in mouse models of Alzheimer's disease (AD) and amyotrophic lateral sclerosis (ALS).⁸⁹ In cell-based models of these conditions, increased SIRT1 promotes neuronal survival.⁸⁹ In animal

models of AD, cortical SIRT1 reduction parallels the accumulation of tau.⁹⁰ In humans with Alzheimer's disease, SIRT1 levels are also reportedly decreased in the parietal cortex but not in the cerebellum. Lower cortical SIRT1 was correlated with the duration of symptoms, lower global cognition scores, and accumulation of amyloid-beta and tau in the cerebral cortex.⁹⁰

SIRT2, the most predominantly expressed sirtuin in the human brain,⁹¹ is enriched in brain oligodendrocytes, where it is thought to be involved in differentiation, maturation, and remodeling.^{92,93} SIRT2 is also highly expressed in post-mitotic neurons and glial cells.⁹⁴ In the brain and other tissues, SIRT2 acts as a tubulin deacetylase,⁹⁵⁻⁹⁷ which inhibits growth in postmitotic neurons⁹⁷ and helps protect neuronal cells against mitotic stress.⁹⁸ SIRT2 is also highly expressed in the myelin sheath, where alpha-tubulin is its main protein target. Decreasing expression of SIRT2 in myelin increases alpha-tubulin acetylation and myelin basic protein expression; increasing expression of SIRT2 has the opposite effect.⁹⁹ Under some experimental circumstances SIRT2 inhibition appears to be neuroprotective.¹⁰⁰ Inhibition of SIRT2 activity also protects against dopaminergic cell death *in vitro* and in a *Drosophila* model of Parkinson's disease.¹⁰¹ Under other circumstances it might be advantageous to express SIRT2. For example, SIRT2 is reportedly reduced in

Figure 3. Biphasic Effect of SIRT1 Overexpression

Protection against oxidative stress, age-dependent increases in cardiac hypertrophy, apoptosis/fibrosis, cardiac dysfunction, and expression of senescence markers



Evidence from transgenic mice bred to overexpress cardiomyocyte SIRT1 suggests a biphasic response when it comes to protecting the heart, with the best response occurring at a level of SIRT1 expression greater than normal but less than the highest levels of expression.

some human brain tumor cell lines, which apparently causes a relative loss of tumor suppressor activity via its role in protein deacetylation.⁹¹

Secondary to its role as a mediator of sirtuin activity, resveratrol appears to have a variety of brain and neuron effects. *In vitro*, by upregulating SIRT1, resveratrol protects neurons from apoptosis by excitotoxins (glutamate and NMDA).¹⁰² Resveratrol improves neuronal cell survival in response to oxidative stress¹⁰³ and protects neuronal cells from ischemic insults.¹⁰⁴ Resveratrol pretreatment of mice is neuroprotective and induces tolerance against brain injury caused by cardiac arrest. These protective effects are associated with a resveratrol-induced increase in hippocampal SIRT1 activity.¹⁰⁵ Existing evidence suggests that resveratrol might counter some aspects of Alzheimer's disease. In cell-based models of AD, SIRT1 is increased and promotes neuronal survival; treatment with resveratrol also promotes survival. In the inducible p25 transgenic mouse, a model of AD and tauopathies, resveratrol reduced neurodegeneration in the hippocampus and prevented learning impairment. Resveratrol also decreased the acetylation of the SIRT1 protein substrates PGC-1 α and p53, which suggests a supportive role on SIRT1 deacetylation.⁸⁹

Resveratrol might be advantageous under some circumstances, but not in others. In cultured cerebellar granule cells taken from slow Wallerian degeneration mice (mice that have delayed axonal degeneration after injury), resveratrol diminished resistance to axonal degeneration. This appeared to occur because resveratrol enhanced neuronal SIRT2, which then promoted tubulin deacetylation that led to axonal degeneration.¹⁰⁶ It appears there might be circumstances where resveratrol would, secondary to its impact on the sirtuin system, result in unwanted responses in the brain nervous system.

Evidence suggests melatonin influences SIRT1. *In vitro*, it acts as a SIRT1 inducer in young and aged neurons¹⁰⁷ and increases SIRT1 and improves deacetylation in senescence-accelerated mice.¹⁴ Limited evidence suggests it might also play a role during sleep deprivation. Rats subjected to total sleep deprivation for five days had reduced SIRT1 activity in hippocampal pyramidal and granular cell layers, which significantly impaired performance on behavioral memory tests. Supplying melatonin preserved SIRT1 activity and resulted in considerably better performance in the memory tests.¹⁰⁸

As was discussed in detail in Part 1 of this review,

nicotinamide is capable of sustaining sirtuin activity (by being recycled into NAD⁺ via its salvage pathway) or inhibiting it, depending on the context. *In vitro* experiments indicate that supplying exogenous nicotinamide preserves NAD⁺ levels, while preventing the excitotoxin-induced reduction in neuron SIRT1 activity.^{86,102} Degeneration of an axon after it is severed can be significantly slowed in the presence of NAD⁺ or its precursors – an effect that appears to be secondary to SIRT1 activation.¹⁰⁹

Because the nicotinamide salvage pathway in the brain is not as robust as in other tissues, the brain might be particularly susceptible to NAD⁺ depletion under circumstances where its rate of use is increased. Supplying nicotinamide under these circumstances appears to regenerate NAD⁺. Evidence suggests that exogenous nicotinamide might act as a sirtuin inhibitor in other circumstances. In AD transgenic mice, oral administration of nicotinamide restored cognitive deficits associated with AD by selectively reducing a specific phospho-species of tau (Thr231) that is associated with microtubule depolymerization, in a manner similar to inhibition of SIRT1. Nicotinamide also dramatically increased acetylated alpha-tubulin, a primary substrate of SIRT2 deacetylase.¹¹⁰ In this study, nicotinamide appeared to inhibit SIRT1 and SIRT2 deacetylation reactions.

Cancer

The current understanding of the relationship between cancer and sirtuins was accurately stated in the title of a review article by Deng – “SIRT1, is it a tumor promoter or tumor suppressor?”¹¹¹ This title aptly captures the current confusion regarding cancer and sirtuins. Deng reports some evidence suggests SIRT1 is a tumor promoter, including increased SIRT1 expression in some cancers,¹¹²⁻¹¹⁸ and its role in deacetylating (and hence presumably deactivating) proteins like p53, p300, and foxhead transcription factors that are involved in tumor suppression and DNA repair.¹¹⁹⁻¹²⁵ Conversely, other cancers have decreased expression of SIRT1.^{117,126-128} Other indications of SIRT1 as a tumor suppressor come from experimental results of mouse/cancer models in which SIRT1 is intentionally under- (tumorigenesis increases) or overexpressed (tumorigenesis is attenuated).^{117,126} SIRT1 also exerts a positive influence on other proteins and processes that result in suppression of tumor growth and enhanced DNA repair.^{126,129-131} Consult the Deng article for an in-depth review.¹¹¹

Like SIRT1, SIRT3 also appears to have both tumor promotion and tumor suppression effects. Although it is capable of deacetylating p53,¹²⁰ it is involved in supporting pro-apoptotic processes by targeting other proteins for deacetylation^{32,132} and functions as a tumor suppressor by enhancing the expression of mitochondrial antioxidant enzymes.¹³³ Mice lacking SIRT3 express genomic instability and develop tumors.¹³³

Conflicting evidence exists, even within the same cancer tissue type. Ashraf et al reported an association between increased SIRT3 and node-positive breast cancer,¹³⁴ while Kim et al reported reduced SIRT3 levels in breast (and other cancers) and noted that mice lacking SIRT3 develop mammary tumors.¹³³

Although less is known about the other sirtuins and cancer, several have functions that suggest a role in cancer prevention. SIRT5 appears to regulate DNA repair and influences apoptosis.¹³⁵ SIRT6 is involved in regulating chromatin structure, maintaining telomere integrity and genomic stability, and repairing DNA.¹³⁶⁻¹⁴¹ SIRT7 promotes ribosomal gene (rDNA) transcription factors and has anti-proliferative effects.^{142,143}

Sirtuin expression is thought to be a protective response to certain forms of stress and toxicity. Some cancer therapies, including radiation and certain forms of chemotherapy, are genotoxic. Limited experimental evidence suggests that the sirtuin system might respond to these treatments to protect cells against them, which might also potentially interfere with the clinical efficacy of these treatments. For example, exposure of cells to radiation caused an increase in SIRT1 and a corresponding increase in DNA repair. Experimentally-induced overexpression of SIRT1 resulted in a greater increase in repair of DNA strand breakages produced by the radiation. Conversely, inhibiting SIRT1 expression resulted in a decrease of DNA repair in response to radiation.¹⁴⁴ Other *in vitro* evidence reported inhibition of SIRT1 expression increased the efficacy of radiation against human lung cancer cells¹⁴⁵ and lack of SIRT1 increased cell sensitivity to radiation.¹⁴⁶ The relationship between SIRT1 and cisplatin has also been investigated *in vitro*. SIRT1 appears to be part of the cellular response to cisplatin, with greater SIRT1 expression associated with increased resistance of cancer cells to this treatment. Conversely, interfering with SIRT1 expression sensitized cells to cisplatin.¹⁴⁷ SIRT1- and SIRT2-deficient cells were also reportedly more sensitive to the pro-apoptotic effects of cisplatin

and staurosporine.¹⁴⁶ This evidence, although *in vitro* and limited, suggests there might be interactions with the sirtuin response and certain cancer therapies that might interfere with or mitigate the efficacy of these therapies.

Resveratrol might have some sirtuin-mediated interactions with cancer. *In vitro* and *in vivo*, SIRT1 appears to be a potential interface between the tumor suppressor gene breast cancer 1 (BRCA1) and survivin (a negative regulator of apoptosis). Experimentally, BRCA1 binds to the SIRT1 gene and increases its expression; SIRT1 in turn inhibits survivin, resulting in programmed cell death. Absence of SIRT1 results in overexpression of survivin and impedes apoptosis. *In vitro*, resveratrol activates SIRT1, which then inhibits survivin expression and promotes apoptosis.¹²⁸ *In vitro*, resveratrol was also a potent sensitizer for cancer drug-induced apoptosis. One of the mechanisms of action for this effect is a downregulation of survivin expression.¹⁴⁸ While this study did not attempt to monitor SIRT1, it is possible that SIRT1 activation was involved in the downregulation of survivin, since SIRT1 is involved in regulating the expression of the survivin gene. *In vitro*, resveratrol dose-dependently induced apoptosis in osteosarcoma cells, but had a minor effect on normal osteoblasts. This difference in effect might be partly explained by SIRT1 expression, since SIRT1 is expressed in higher amounts in osteosarcoma cell lines than in normal human osteoblasts.¹⁴⁹ *In vitro*, resveratrol promotes autophagy (a mechanism that causes death of stressed cells by means other than apoptotic or necrotic demise), apparently mediated by SIRT1 activation.^{150,151} Resveratrol's activation of SIRT1 also promoted improved DNA repair activity subsequent to genotoxic stress.¹⁵² Mice bred to underexpress SIRT1 and p53 develop tumors in multiple tissues, and administration of resveratrol reduced tumorigenesis in these mice.¹²⁷ Topical application of resveratrol has been reported to reduce tumorigenesis in a mouse model of skin cancer, an effect that was significantly reduced in mice lacking the SIRT1 gene.¹⁵³

Conclusion

As research has better characterized the sirtuin system, it has become apparent that this system regulates many proteins, which themselves influence a variety of cellular processes. Because of their impact on the function of a diverse array of proteins, sirtuins are involved with metabolic responses and processes that influence many aspects of human function. Existing evidence

strongly supports sirtuin involvement in longevity, age-related diseases, obesity, cardiovascular and neurological function, and cancer.

As the responses become better understood, which sirtuins to target for activation or inhibition should become clearer. Cancer is a good example. Experimental evidence argues that sirtuins play a complex, more nuanced role in cancer than can be determined by its effects on any protein or metabolic process viewed in isolation. The complicated and perhaps competing effects of individual sirtuins on cellular processes that influence cancer development, suppression, and progression suggest much more research is required. Although SIRT1 has been found to increase in some cancers and not in others, its increase alone cannot be taken as evidence that it is a cause of cancer development. On the other hand, it could be a *consequence* of tumorigenesis or other factors involved in cancer or an adaptive response intended to counter genotoxic insults that contribute to cancer. Although sirtuin expression might counteract the desired clinical response to certain cancer therapies, specifically radiation and chemotherapy, there might be times when an increased sirtuin response might enhance cancer prevention or treatment. Currently there are as many questions as there are answers.

Resveratrol has generally been characterized as a sirtuin activator. It is possible that this might be an oversimplification of its actions. While it does appear to activate sirtuins under most circumstances, some evidence suggests a more adaptive effect on sirtuins. Available *in vitro* and *in vivo* evidence suggests that resveratrol is most likely to produce a noticeable physiological effect under stressful circumstances or those involving unhealthy lifestyle habits. For example, when mice were fed standard- and high-fat rat chow diets, the effects of resveratrol were significantly more dramatic in countering the effects of the latter diet. Presumably, this is because expression and activity of sirtuins are strongly influenced by environmental factors, especially dietary, lifestyle, or environment factors that create some form of stress. Although resveratrol might play a significant role in augmenting the sirtuin response, human research is required before any definitive inferences can be made.

References

- Vinciguerra M, Fulco M, Ladurner A, et al. SirT1 in muscle physiology and disease: lessons from mouse models. *Dis Model Mech* 2010;3:298-303.
- Li Y, Xu W, McBurney MW, Longo VD. SirT1 inhibition reduces IGF-1/IRS-2/Ras/ERK1/2 signaling and protects neurons. *Cell Metab* 2008;8:38-48.
- Howitz KT, Bitterman KJ, Cohen HY, et al. Small molecule activators of sirtuins extend *Saccharomyces cerevisiae* lifespan. *Nature* 2003;425:191-196.
- Kaeberlein M, McDonagh T, Heltweg B, et al. Substrate-specific activation of sirtuins by resveratrol. *J Biol Chem* 2005;280:17038-17045.
- Wood JG, Rogina B, Lavu S, et al. Sirtuin activators mimic caloric restriction and delay ageing in metazoans. *Nature* 2004;430:686-689.
- Bass TM, Weinkove D, Houthoofd K, et al. Effects of resveratrol on lifespan in *Drosophila melanogaster* and *Caenorhabditis elegans*. *Mech Ageing Dev* 2007;128:546-552.
- Zou S, Carey JR, Liedo P, et al. The prolongevity effect of resveratrol depends on dietary composition and calorie intake in a tephritid fruit fly. *Exp Gerontol* 2009;44:472-476.
- Valenzano DR, Terzibasi E, Genade T, et al. Resveratrol prolongs lifespan and retards the onset of age-related markers in a short-lived vertebrate. *Curr Biol* 2006;16:296-300.
- Baur JA, Pearson KJ, Price NL, et al. Resveratrol improves health and survival of mice on a high-calorie diet. *Nature* 2006;444:337-342.
- Barger JL, Kayo T, Vann JM, et al. A low dose of dietary resveratrol partially mimics caloric restriction and retards aging parameters in mice. *PLoS One* 2008;3:e2264.
- Pearson KJ, Baur JA, Lewis KN, et al. Resveratrol delays age-related deterioration and mimics transcriptional aspects of dietary restriction without extending life span. *Cell Metab* 2008;8:157-168.
- Yokozawa T, Lee YA, Zhao Q, et al. Persimmon oligomeric proanthocyanidins extend life span of senescence-accelerated mice. *J Med Food* 2009;12:1199-1205.
- Pietsch K, Saul N, Menzel R, et al. Quercetin mediated lifespan extension in *Caenorhabditis elegans* is modulated by age-1, daf-2, sek-1 and unc-43. *Biogerontology* 2009;10:565-578.
- Gutierrez-Cuesta J, Tajés M, Jiménez A, et al. Evaluation of potential pro-survival pathways regulated by melatonin in a murine senescence model. *J Pineal Res* 2008;45:497-505.
- Liang F, Kume S, Koya D. SIRT1 and insulin resistance. *Nat Rev Endocrinol* 2009;5:367-373.
- Yoshizaki T, Milne JC, Imamura T, et al. SIRT1 exerts anti-inflammatory effects and improves insulin sensitivity in adipocytes. *Mol Cell Biol* 2009;29:1363-1374.

17. Qiao L, Shao J. SIRT1 regulates adiponectin gene expression through Foxo1-C/EBPalpha transcriptional complex. *J Biol Chem* 2006;281:39915-39924.
18. Wang H, Qiang L, Farmer SR. Identification of a domain within peroxisome proliferator-activated receptor gamma regulating expression of a group of genes containing fibroblast growth factor 21 that are selectively repressed by SIRT1 in adipocytes. *Mol Cell Biol* 2008;28:188-200.
19. Cakir I, Perello M, Lansari O, et al. Hypothalamic Sirt1 regulates food intake in a rodent model system. *PLoS One* 2009;4:e8322.
20. Picard F, Kurtev M, Chung N, et al. Sirt1 promotes fat mobilization in white adipocytes by repressing PPAR-gamma. *Nature* 2004;429:771-776.
21. Ramadori G, Lee CE, Bookout AL, et al. Brain SIRT1: anatomical distribution and regulation by energy availability. *J Neurosci* 2008;28:9989-9996.
22. Chen D, Steele AD, Lindquist S, Guarente L. Increase in activity during calorie restriction requires Sirt1. *Science* 2005;310:1641.
23. Banks AS, Kon N, Knight C, et al. SirT1 gain of function increases energy efficiency and prevents diabetes in mice. *Cell Metab* 2008;8:333-341.
24. Bordone L, Cohen D, Robinson et al. SIRT1 transgenic mice show phenotypes resembling calorie restriction. *Aging Cell* 2007;6:759-767.
25. Pfluger PT, Herranz D, Velasco-Miguel S, et al. Sirt1 protects against high-fat diet-induced metabolic damage. *Proc Natl Acad Sci U S A* 2008;105:9793-9798.
26. Sun C, Zhang F, Ge X, et al. SIRT1 improves insulin sensitivity under insulin-resistant conditions by repressing PTP1B. *Cell Metab* 2007;6:307-319.
27. Bordone L, Motta MC, Picard F, et al. Sirt1 regulates insulin secretion by repressing UCP2 in pancreatic beta cells. *PLoS Biol* 2006;4:e31.
28. Moynihan KA, Grimm AA, Plueger MM, et al. Increased dosage of mammalian Sir2 in pancreatic beta cells enhances glucose-stimulated insulin secretion in mice. *Cell Metab* 2005;2:105-117.
29. Ramsey KM, Mills KF, Satoh A, Imai S. Age-associated loss of Sirt1-mediated enhancement of glucose-stimulated insulin secretion in beta cell-specific Sirt1-overexpressing (BESTO) mice. *Aging Cell* 2008;7:78-88.
30. Li X, Zhang S, Blander G, et al. SIRT1 deacetylates and positively regulates the nuclear receptor LXR. *Mol Cell* 2007;28:91-106.
31. Jing E, Gesta S, Kahn CR. SIRT2 regulates adipocyte differentiation through FoxO1 acetylation/deacetylation. *Cell Metab* 2007;6:105-114.
32. Hirschev MD, Shimazu T, Goetzman E, et al. SIRT3 regulates mitochondrial fatty-acid oxidation by reversible enzyme deacetylation. *Nature* 2010;464:121-125.
33. Shi T, Wang F, Stieren E, Tong Q. SIRT3, a mitochondrial sirtuin deacetylase, regulates mitochondrial function and thermogenesis in brown adipocytes. *J Biol Chem* 2005;280:13560-13567.
34. Ahuja N, Schwer B, Carobbio S, et al. Regulation of insulin secretion by SIRT4, a mitochondrial ADP-ribosyltransferase. *J Biol Chem* 2007;282:33583-33592.
35. Zhong L, D'Urso A, Toiber D, et al. The histone deacetylase Sirt6 regulates glucose homeostasis via Hif1alpha. *Cell* 2010;140:280-293.
36. Kanfi Y, Peshti V, Gil R, et al. SIRT6 protects against pathological damage caused by diet-induced obesity. *Aging Cell* 2010;9:162-173.
37. Pedersen SB, Ølholm J, Paulsen SK, et al. Low Sirt1 expression, which is upregulated by fasting, in human adipose tissue from obese women. *Int J Obes (Lond)* 2008;32:1250-1255.
38. Rutanen J, Yaluri N, Modi S, et al. SIRT1 mRNA expression may be associated with energy expenditure and insulin sensitivity. *Diabetes* 2010;59:829-835.
39. de Kreutzenberg SV, Ceolotto G, Papparella I, et al. Downregulation of the longevity-associated protein sirtuin 1 in insulin resistance and metabolic syndrome: potential biochemical mechanisms. *Diabetes* 2010;59:1006-1015.
40. Heilbronn LK, Civitarese AE, Bogacka I, et al. Glucose tolerance and skeletal muscle gene expression in response to alternate day fasting. *Obes Res* 2005;13:574-581.
41. Capel F, Viguier N, Vega N, et al. Contribution of energy restriction and macronutrient composition to changes in adipose tissue gene expression during dietary weight-loss programs in obese women. *J Clin Endocrinol Metab* 2008;93:4315-4322.
42. Bai L, Pang WJ, Yang YJ, Yang GS. Modulation of Sirt1 by resveratrol and nicotinamide alters proliferation and differentiation of pig preadipocytes. *Mol Cell Biochem* 2008;307:129-140.
43. Shan T, Wang Y, Wu T, et al. Porcine sirtuin 1 gene clone, expression pattern, and regulation by resveratrol. *J Anim Sci* 2009;87:895-904.
44. Wang GL, Fu YC, Xu WC, et al. Resveratrol inhibits the expression of SREBP1 in cell model of steatosis via Sirt1-FOXO1 signaling pathway. *Biochem Biophys Res Commun* 2009;380:644-649.
45. Bäckesjö CM, Li Y, Lindgren U, Haldosén LA. Activation of Sirt1 decreases adipocyte formation during osteoblast differentiation of mesenchymal stem cells. *J Bone Miner Res* 2006;21:993-1002.
46. Breen DM, Sanli T, Giacca A, Tsiani E. Stimulation of muscle cell glucose uptake by resveratrol through sirtuins and AMPK. *Biochem Biophys Res Commun* 2008;374:117-122.
47. Lee JH, Song MY, Song EK, et al. Overexpression of SIRT1 protects pancreatic beta-cells against cytokine toxicity by suppressing the nuclear factor-kappaB signaling pathway. *Diabetes* 2009;58:344-351.
48. Um JH, Park SJ, Kang H, et al. AMP-activated protein kinase-deficient mice are resistant to the metabolic effects of resveratrol. *Diabetes* 2010;59:554-563.
49. Purushotham A, Schug TT, Li X. SIRT1 performs a balancing act on the tight-rope toward longevity. *Aging (Albany NY)* 2009;1:669-673.
50. Xu F, Gao Z, Zhang J, et al. Lack of SIRT1 (Mammalian Sirtuin 1) activity leads to liver steatosis in the Sirt1+/- mice: a role of lipid mobilization and inflammation. *Endocrinology* 2010. [Epub ahead of print]
51. Escande C, Chini CC, Nin V, et al. Deleted in breast cancer-1 regulates SIRT1 activity and contributes to high-fat diet induced liver steatosis in mice. *J Clin Invest* 2010;120:545-558.

52. Deng XQ, Chen LL, Li NX. The expression of SIRT1 in nonalcoholic fatty liver disease induced by high-fat diet in rats. *Liver Int* 2007;27:708-715.
53. Yamazaki Y, Usui I, Kanatani Y, et al. Treatment with SIRT1720, a SIRT1 activator, ameliorates fatty liver with reduced expression of lipogenic enzymes in MSG mice. *Am J Physiol Endocrinol Metab* 2009. [Epub ahead of print]
54. Hou X, Xu S, Maitland-Toolan KA, et al. SIRT1 regulates hepatocyte lipid metabolism through activating AMP-activated protein kinase. *J Biol Chem* 2008;283:20015-20026.
55. Ajmo JM, Liang X, Rogers CQ, et al. Resveratrol alleviates alcoholic fatty liver in mice. *Am J Physiol Gastrointest Liver Physiol* 2008;295:G833-G842.
56. Lieber CS, Leo MA, Wang X, Decarli LM. Effect of chronic alcohol consumption on hepatic SIRT1 and PGC-1alpha in rats. *Biochem Biophys Res Commun* 2008;370:44-48.
57. Oliva J, French BA, Li J, et al. Sirt1 is involved in energy metabolism: the role of chronic ethanol feeding and resveratrol. *Exp Mol Pathol* 2008;85:155-159.
58. You M, Cao Q, Liang X, et al. Mammalian sirtuin 1 is involved in the protective action of dietary saturated fat against alcoholic fatty liver in mice. *J Nutr* 2008;138:497-501.
59. You M, Liang X, Ajmo JM, Ness GC. Involvement of mammalian sirtuin 1 in the action of ethanol in the liver. *Am J Physiol Gastrointest Liver Physiol* 2008;294:G892-G898.
60. Shi T, Fan GQ, Xiao SD. SIRT3 reduces lipid accumulation via AMPK activation in human hepatic cells. *J Dig Dis* 2010;11:55-62.
61. Palacios OM, Carmona JJ, Michan S, et al. Diet and exercise signals regulate SIRT3 and activate AMPK and PGC-1alpha in skeletal muscle. *Aging (Albany NY)* 2009;1:771-783.
62. Costa Cdos S, Hammes TO, Rohden F, et al. SIRT1 transcription is decreased in visceral adipose tissue of morbidly obese patients with severe hepatic steatosis. *Obes Surg* 2010;20:633-639.
63. Balestrieri ML, Rienzo M, Felice F, et al. High glucose downregulates endothelial progenitor cell number via SIRT1. *Biochim Biophys Acta* 2008;1784:936-945.
64. Potente M, Ghaeni L, Baldessari D, et al. SIRT1 controls endothelial angiogenic functions during vascular growth. *Genes Dev* 2007;21:2644-2658.
65. Mattagajasingh I, Kim CS, Naqvi A, et al. SIRT1 promotes endothelium-dependent vascular relaxation by activating endothelial nitric oxide synthase. *Proc Natl Acad Sci U S A* 2007;104:14855-14860.
66. Orimo M, Minamino T, Miyauchi H, et al. Protective role of SIRT1 in diabetic vascular dysfunction. *Arterioscler Thromb Vasc Biol* 2009;29:889-894.
67. Arunachalam G, Yao H, Sundar IK, et al. SIRT1 regulates oxidant- and cigarette smoke-induced eNOS acetylation in endothelial cells: role of resveratrol. *Biochem Biophys Res Commun* 2010;393:66-72.
68. Gracia-Sancho J, Villarreal G Jr, Zhang Y, Garcia-Cardena G. Activation of SIRT1 by resveratrol induces KLF2 expression conferring an endothelial vasoprotective phenotype. *Cardiovasc Res* 2010;85:514-519.
69. Miyazaki R, Ichiki T, Hashimoto T, et al. SIRT1, a longevity gene, downregulates angiotensin II type 1 receptor expression in vascular smooth muscle cells. *Arterioscler Thromb Vasc Biol* 2008;28:1263-1269.
70. Cardellini M, Menghini R, Martelli E, et al. TIMP3 is reduced in atherosclerotic plaques from subjects with type 2 diabetes and increased by Sirt1. *Diabetes* 2009;58:2396-2401.
71. Sundaresan NR, Samant SA, Pillai VB, et al. SIRT3 is a stress-responsive deacetylase in cardiomyocytes that protects cells from stress-mediated cell death by deacetylation of Ku70. *Mol Cell Biol* 2008;28:6384-6401.
72. Rajamohan SB, Pillai VB, Gupta M, et al. SIRT1 promotes cell survival under stress by deacetylation-dependent deactivation of poly(ADP-ribose) polymerase 1. *Mol Cell Biol* 2009;29:4116-4129.
73. Tanno M, Kuno A, Yano T, et al. Induction of manganese superoxide dismutase by nuclear translocation and activation of SIRT1 promotes cell survival in chronic heart failure. *J Biol Chem* 2010;285:8375-8382.
74. Sundaresan NR, Gupta M, Kim G, et al. Sirt3 blocks the cardiac hypertrophic response by augmenting Foxo3a-dependent antioxidant defense mechanisms in mice. *J Clin Invest* 2009;119:2758-2771.
75. Sulaiman M, Matta MJ, Sunderesan NR, et al. Resveratrol, an activator of SIRT1, upregulates sarcoplasmic calcium ATPase and improves cardiac function in diabetic cardiomyopathy. *Am J Physiol Heart Circ Physiol* 2010;298:H833-H843.
76. Chen CJ, Yu W, Fu YC, et al. Resveratrol protects cardiomyocytes from hypoxia-induced apoptosis through the SIRT1-FoxO1 pathway. *Biochem Biophys Res Commun* 2009;378:389-393.
77. Danz ED, Skramsted J, Henry N, et al. Resveratrol prevents doxorubicin cardiotoxicity through mitochondrial stabilization and the Sirt1 pathway. *Free Radic Biol Med* 2009;46:1589-1597.
78. Mukherjee S, Lekli I, Gurusamy N, et al. Expression of the longevity proteins by both red and white wines and their cardioprotective components, resveratrol, tyrosol, and hydroxytyrosol. *Free Radic Biol Med* 2009;46:573-578.
79. Yoshida Y, Shioi T, Izumi T. Resveratrol ameliorates experimental autoimmune myocarditis. *Circ J* 2007;71:397-404.
80. Alcendor RR, Gao S, Zhai P, et al. Sirt1 regulates aging and resistance to oxidative stress in the heart. *Circ Res* 2007;100:1512-1521.
81. Ahn BH, Kim HS, Song S, et al. A role for the mitochondrial deacetylase Sirt3 in regulating energy homeostasis. *Proc Natl Acad Sci U S A* 2008;105:14447-14452.
82. Vakhrusheva O, Smolka C, Gajawada P, et al. Sirt7 increases stress resistance of cardiomyocytes and prevents apoptosis and inflammatory cardiomyopathy in mice. *Circ Res* 2008;102:703-710.
83. Pfister JA, Ma C, Morrison BE, D'Mello SR. Opposing effects of sirtuins on neuronal survival: SIRT1-mediated neuroprotection is independent of its deacetylase activity. *PLoS One* 2008;3:e4090.
84. Zakhary SM, Ayubcha D, Dileo JN, et al. Distribution analysis of deacetylase SIRT1 in rodent and human nervous systems. *Anat Rec (Hoboken)* 2010. [Epub ahead of print]
85. Chen D, Steele AD, Hutter G, et al. The role of calorie restriction and SIRT1 in prion-mediated neurodegeneration. *Exp Gerontol* 2008;43:1086-1093.

86. Liu D, Pitta M, Mattson MP. Preventing NAD(+) depletion protects neurons against excitotoxicity: bioenergetic effects of mild mitochondrial uncoupling and caloric restriction. *Ann N Y Acad Sci* 2008;1147:275-282.
87. Qin W, Yang T, Ho L, et al. Neuronal SIRT1 activation as a novel mechanism underlying the prevention of Alzheimer disease amyloid neuropathology by calorie restriction. *J Biol Chem* 2006;281:21745-21754.
88. Kakefuda K, Fujita Y, Oyagi A, et al. Sirtuin 1 overexpression mice show a reference memory deficit, but not neuroprotection. *Biochem Biophys Res Commun* 2009;387:784-788.
89. Kim D, Nguyen MD, Dobbin MM, et al. SIRT1 deacetylase protects against neurodegeneration in models for Alzheimer's disease and amyotrophic lateral sclerosis. *EMBO J* 2007;26:3169-3179.
90. Julien C, Tremblay C, Emond V, et al. Sirtuin 1 reduction parallels the accumulation of tau in Alzheimer disease. *J Neuropathol Exp Neurol* 2009;68:48-58.
91. Voelter-Mahlknecht S, Ho AD, Mahlknecht U. FISH-mapping and genomic organization of the NAD-dependent histone deacetylase gene, Sirtuin 2 (Sirt2). *Int J Oncol* 2005;27:1187-1196.
92. Tang BL, Chua CE. SIRT2, tubulin deacetylation, and oligodendroglia differentiation. *Cell Motil Cytoskeleton* 2008;65:179-182.
93. Southwood CM, Peppi M, Dryden S, et al. Microtubule deacetylases, SirT2 and HDAC6, in the nervous system. *Neurochem Res* 2007;32:187-195.
94. Harting K, Knöll B. SIRT2-mediated protein deacetylation: an emerging key regulator in brain physiology and pathology. *Eur J Cell Biol* 2010;89:262-269.
95. Peck B, Chen CY, Ho KK, et al. SIRT inhibitors induce cell death and p53 acetylation through targeting both SIRT1 and SIRT2. *Mol Cancer Ther* 2010;9:844-855.
96. North BJ, Marshall BL, Borra MT, et al. The human Sir2 ortholog, SIRT2, is an NAD+-dependent tubulin deacetylase. *Mol Cell* 2003;11:437-444.
97. Pandithage R, Lilischkis R, Harting K, et al. The regulation of SIRT2 function by cyclin-dependent kinases affects cell motility. *J Cell Biol* 2008;180:915-929.
98. Inoue T, Hiratsuka M, Osaki M, et al. SIRT2, a tubulin deacetylase, acts to block the entry to chromosome condensation in response to mitotic stress. *Oncogene* 2007;26:945-957.
99. Li W, Zhang B, Tang J, et al. Sirtuin 2, a mammalian homolog of yeast silent information regulator-2 longevity regulator, is an oligodendroglial protein that decelerates cell differentiation through deacetylating alpha-tubulin. *J Neurosci* 2007;27:2606-2616.
100. Luthi-Carter R, Taylor DM, Pallos J, et al. SIRT2 inhibition achieves neuroprotection by decreasing sterol biosynthesis. *Proc Natl Acad Sci U S A* 2010;107:7927-7932.
101. Outeiro TF, Kontopoulos E, Altmann SM, et al. Sirtuin 2 inhibitors rescue alpha-synuclein-mediated toxicity in models of Parkinson's disease. *Science* 2007;317:516-519.
102. Liu D, Gharavi G, Pitta M, et al. Nicotinamide prevents NAD+ depletion and protects neurons against excitotoxicity and cerebral ischemia: NAD+ consumption by SIRT1 may endanger energetically compromised neurons. *Neuromolecular Med* 2009;11:28-42.
103. Chong ZZ, Maiese K. Enhanced tolerance against early and late apoptotic oxidative stress in mammalian neurons through nicotinamidase and sirtuin mediated pathways. *Curr Neurovasc Res* 2008;5:159-170.
104. Raval AP, Dave KR, Pérez-Pinzón MA. Resveratrol mimics ischemic preconditioning in the brain. *J Cereb Blood Flow Metab* 2006;26:1141-1147.
105. Della-Morte D, Dave KR, DeFazio RA, et al. Resveratrol pretreatment protects rat brain from cerebral ischemic damage via a sirtuin 1-uncoupling protein 2 pathway. *Neuroscience* 2009;159:993-1002.
106. Suzuki K, Koike T. Resveratrol abolishes resistance to axonal degeneration in slow Wallerian degeneration (Wlds) mice: activation of SIRT2, an NAD-dependent tubulin deacetylase. *Biochem Biophys Res Commun* 2007;359:665-671.
107. Tajés M, Gutierrez-Cuesta J, Ortuño-Sahagun D, et al. Anti-aging properties of melatonin in an *in vitro* murine senescence model: involvement of the sirtuin 1 pathway. *J Pineal Res* 2009;47:228-237.
108. Chang HM, Wu UI, Lan CT. Melatonin preserves longevity protein (sirtuin 1) expression in the hippocampus of total sleep-deprived rats. *J Pineal Res* 2009;47:211-220.
109. Araki T, Sasaki Y, Milbrandt J. Increased nuclear NAD biosynthesis and SIRT1 activation prevent axonal degeneration. *Science* 2004;305:1010-1013.
110. Green KN, Steffan JS, Martinez-Coria H, et al. Nicotinamide restores cognition in Alzheimer's disease transgenic mice via a mechanism involving sirtuin inhibition and selective reduction of Thr231-phosphotau. *J Neurosci* 2008;28:11500-11510.
111. Deng CX. SIRT1, is it a tumor promoter or tumor suppressor? *Int J Biol Sci* 2009;5:147-152.
112. Bradbury CA, Khanim FL, Hayden R, et al. Histone deacetylases in acute myeloid leukaemia show a distinctive pattern of expression that changes selectively in response to deacetylase inhibitors. *Leukemia* 2005;19:1751-1759.
113. Cha EJ, Noh SJ, Kwon KS, et al. Expression of DBC1 and SIRT1 is associated with poor prognosis of gastric carcinoma. *Clin Cancer Res* 2009;15:4453-4459.
114. Hida Y, Kubo Y, Murao K, Arase S. Strong expression of a longevity-related protein, SIRT1, in Bowen's disease. *Arch Dermatol Res* 2007;299:103-106.
115. Huffman DM, Grizzle WE, Bamman MM, et al. SIRT1 is significantly elevated in mouse and human prostate cancer. *Cancer Res* 2007;67:6612-6618.
116. Jung-Hynes B, Nihal M, Zhong W, Ahmad N. Role of sirtuin histone deacetylase SIRT1 in prostate cancer. A target for prostate cancer management via its inhibition? *J Biol Chem* 2009;284:3823-3832.
117. Kabra N, Li Z, Chen L, et al. SirT1 is an inhibitor of proliferation and tumor formation in colon cancer. *J Biol Chem* 2009;284:18210-18217.
118. Stunkel W, Peh BK, Tan YC, et al. Function of the SIRT1 protein deacetylase in cancer. *Biotechnol J* 2007;2:1360-1368.
119. Brunet A, Sweeney LB, Sturgill JF, et al. Stress-dependent regulation of FOXO transcription factors by the SIRT1 deacetylase. *Science* 2004;303:2011-2015.

120. Li S, Banck M, Mujtaba S, et al. p53-Induced growth arrest is regulated by the mitochondrial SirT3 deacetylase. *PLoS One* 2010;5:e10486.
121. Luo J, Nikolaev AY, Imai S, et al. Negative control of p53 by Sir2alpha promotes cell survival under stress. *Cell* 2001;107:137-148.
122. Vaziri H, Dessain SK, Ng Eaton E, et al. hSIR2(SIRT1) functions as an NAD-dependent p53 deacetylase. *Cell* 2001;107:149-159.
123. Jung-Hynes B, Ahmad N. Role of p53 in the anti-proliferative effects of Sirt1 inhibition in prostate cancer cells. *Cell Cycle* 2009;8:1478-1483.
124. Yamakuchi M, Lowenstein CJ. MiR-34, SIRT1 and p53: the feedback loop. *Cell Cycle* 2009;8:712-715.
125. Zhao W, Kruse JP, Tang Y, et al. Negative regulation of the deacetylase SIRT1 by DBC1. *Nature* 2008;451:587-590.
126. Firestein R, Blander G, Michan S, et al. The SIRT1 deacetylase suppresses intestinal tumorigenesis and colon cancer growth. *PLoS One* 2008;3:e2020.
127. Wang RH, Sengupta K, Li C, et al. Impaired DNA damage response, genome instability, and tumorigenesis in SIRT1 mutant mice. *Cancer Cell* 2008;14:312-323.
128. Wang RH, Zheng Y, Kim HS, et al. Interplay among BRCA1, SIRT1, and Survivin during BRCA1-associated tumorigenesis. *Mol Cell* 2008;32:11-20.
129. Chua KF, Mostoslavsky R, Lombard DB, et al. Mammalian SIRT1 limits replicative life span in response to chronic genotoxic stress. *Cell Metab* 2005;2:67-76.
130. Oberdoerffer P, Michan S, McVay M, et al. SIRT1 redistribution on chromatin promotes genomic stability but alters gene expression during aging. *Cell* 2008;135:907-918.
131. Pruitt K, Zinn RL, Ohm JE, et al. Inhibition of SIRT1 reactivates silenced cancer genes without loss of promoter DNA hypermethylation. *PLoS Genet* 2006;2:e40.
132. Allison SJ, Milner J. SIRT3 is pro-apoptotic and participates in distinct basal apoptotic pathways. *Cell Cycle* 2007;6:2669-2677.
133. Kim HS, Patel K, Muldoon-Jacobs K, et al. SIRT3 is a mitochondria-localized tumor suppressor required for maintenance of mitochondrial integrity and metabolism during stress. *Cancer Cell* 2010;17:41-52.
134. Ashraf N, Zino S, Macintyre A, et al. Altered sirtuin expression is associated with node-positive breast cancer. *Br J Cancer* 2006;95:1056-1061.
135. Schlicker C, Gertz M, Papatheodorou P, et al. Substrates and regulation mechanisms for the human mitochondrial sirtuins Sirt3 and Sirt5. *J Mol Biol* 2008;382:790-801.
136. Tennen RI, Berber E, Chua KF. Functional dissection of SIRT6: identification of domains that regulate histone deacetylase activity and chromatin localization. *Mech Ageing Dev* 2010;131:185-192.
137. McCord RA, Michishita E, Hong T, et al. SIRT6 stabilizes DNA-dependent protein kinase at chromatin for DNA double-strand break repair. *Aging (Albany NY)* 2009;1:109-121.
138. Kawahara TL, Michishita E, Adler AS, et al. SIRT6 links histone H3 lysine 9 deacetylation to NF-kappaB-dependent gene expression and organismal life span. *Cell* 2009;136:62-74.
139. Lombard DB. Sirtuins at the breaking point: SIRT6 in DNA repair. *Aging (Albany NY)* 2009;1:12-16.
140. Michishita E, McCord RA, Berber E, et al. SIRT6 is a histone H3 lysine 9 deacetylase that modulates telomeric chromatin. *Nature* 2008;452:492-496.
141. Mostoslavsky R, Chua KF, Lombard DB, et al. Genomic instability and aging-like phenotype in the absence of mammalian SIRT6. *Cell* 2006;124:315-329.
142. Grob A, Roussel P, Wright JE, et al. Involvement of SIRT7 in resumption of rDNA transcription at the exit from mitosis. *J Cell Sci* 2009;122:489-498.
143. Vakhrusheva O, Braeuer D, Liu Z, et al. Sirt7-dependent inhibition of cell growth and proliferation might be instrumental to mediate tissue integrity during aging. *J Physiol Pharmacol* 2008;59:201-212.
144. Jeong J, Juhn K, Lee H, et al. SIRT1 promotes DNA repair activity and deacetylation of Ku70. *Exp Mol Med* 2007;39:8-13.
145. Sun Y, Sun D, Li F, et al. Downregulation of SIRT1 by antisense oligonucleotides induces apoptosis and enhances radiation sensitization in A549 lung cancer cells. *Lung Cancer* 2007;58:21-29.
146. Matsushita N, Takami Y, Kimura M, et al. Role of NAD-dependent deacetylases SIRT1 and SIRT2 in radiation and cisplatin-induced cell death in vertebrate cells. *Genes Cells* 2005;10:321-332.
147. Liang XJ, Finkel T, Shen DW, et al. SIRT1 contributes in part to cisplatin resistance in cancer cells by altering mitochondrial metabolism. *Mol Cancer Res* 2008;6:1499-1506.
148. Fulda S, Debatin KM. Sensitization for anticancer drug-induced apoptosis by the chemopreventive agent resveratrol. *Oncogene* 2004;23:6702-6711.
149. Li Y, Bäckesjö CM, Haldosén LA, Lindgren U. Resveratrol inhibits proliferation and promotes apoptosis of osteosarcoma cells. *Eur J Pharmacol* 2009;609:13-18.
150. Morselli E, Galluzzi L, Kepp O, et al. Autophagy mediates pharmacological lifespan extension by spermidine and resveratrol. *Aging (Albany NY)* 2009;1:961-970.
151. Morselli E, Maiuri MC, Markaki M, et al. The life span-prolonging effect of sirtuin-1 is mediated by autophagy. *Autophagy* 2010;6:186-188.
152. Yamamori T, DeRicco J, Naqvi A, et al. SIRT1 deacetylates APE1 and regulates cellular base excision repair. *Nucleic Acids Res* 2010;38:832-845.
153. Boily G, He XH, Pearce B, et al. SIRT1-null mice develop tumors at normal rates but are poorly protected by resveratrol. *Oncogene* 2009;28:2882-2893.